

## Comparative Effects of Some PGRs Combination on Proliferation and Hyperhydricity of Sebri Pear Cultivar

S. Karimpour<sup>1</sup>, G. H. Davarynejad<sup>1</sup>, A. Tehranifar<sup>1</sup> and A. Bagheri<sup>2</sup>

1- Department of Horticultural Science, Ferdowsi University of Mashhad, Mashhad, Iran

2- Department of Biotechnology and plant breeding, Ferdowsi University of Mashhad, Mashhad, Iran

Corresponding author Email: [s.karimpour@yahoo.com](mailto:s.karimpour@yahoo.com)

**ABSTRACT:** In order to improve *in vitro* culture of Sebri pear cultivar several combinations of plant growth regulators of BAP, IBA and GA<sub>3</sub> were tested. In all PGRs combination, regenerated shoots increased with increasing BAP concentration. In media containing of different combinations of BAP (3 mgL<sup>-1</sup>) and BAP (2 mgL<sup>-1</sup>) + GA<sub>3</sub> the highest of regenerated shoots were obtained. The higher regenerated shoot length showed in media containing IBA. The highest regenerated shoots length obtained in BAP (2 mgL<sup>-1</sup>) + IBA and the lowest showed in BAP (3 mgL<sup>-1</sup>). In BAP (3 mgL<sup>-1</sup>) + GA<sub>3</sub> combination, internode length was the highest. Hyperhydricity rate was affected by BAP concentration and plant growth regulators combinations. A negative relation was obtained in internode length and hyperhydricity rate. The higher concentration of BAP induced more hyperhydricity rate and BAP + IBA + GA<sub>3</sub> produced the highest hyperhydric regenerated shoots, while BAP (2 mgL<sup>-1</sup>) + GA<sub>3</sub> induced the lowest hyperhydricity rate.

**Keywords:** BAP, GA<sub>3</sub>, IBA, Micro propagation, Tissue culture

**Abbreviations:** BAP: 6-benzylaminopurine; IBA: Indole-3-butyric acid; GA<sub>3</sub>: Gibberelic acid; MS: Murashige and Skoog medium (1962), PGRs: Plant growth regulators

### INTRODUCTION

Pear is one of the most important fruit trees that widely have been grown in moderate regions such as Iran. It is reported Sebri cultivar is belonged to *P. communis* L. (Safarpour, 2008; Erfani et al., 2012) but some author are mentioned *P. pyrifolia* (Rahemi and Baghbani, 2002; Davarynejad and Davarynejad, 2007; Zafari nia et al., 2010). Micropropagation of Sebri was a strategy for its propagation, because of the difficulty of rooting and also budding or grafting of this cultivar on common rootstock have been incompatible (Amiri, 2002; Davarynejad and Davarynejad, 2007; Davarynejad et al., 2008).

Different concentration and combination of plant growth regulators may be have various effects on proliferation (Pierik, 1990; Grattapagali and Machado, 1998). Many workers have suggested combination of BAP and low concentration of auxins for proliferation of pear species (Yeo and Reed, 1995; Nadosy, 1997; Dwivedi and Bist, 1999; Nosrati et al., 2009). Hyperhydricity

(previously known as vitrification) of regenerated shoots is an undesirable phenomena *in vitro* culture. In pear, hyperhydricity was affected by some factors such as cultivars, type and concentration of cytokinins (Rouzban et al., 2002; Kadota and Niimi, 2003) and medium type (Kadota et al., 2001). The aim of this study was to investigate the influence of BAP concentration and plant growth regulators combinations on regeneration and hyperhydricity rate in proliferation stage of Sebri pear cultivar.

### MATERIALS AND METHODS

*In vitro* shoots of Sebri cultivar were used for this experiment. Shoots were cultured on MS (Murashige and Skoog, 1962) medium supplement with 30 gL<sup>-1</sup> sucrose and different combinations of plant growth regulators whit two BAP concentrations (2 and 3 mgL<sup>-1</sup>) and four combinations of PGRs (BAP, BAP + IBA, BAP + GA<sub>3</sub>,

BAP + IBA + GA<sub>3</sub>) with constant concentration of IBA (0.1 mgL<sup>-1</sup>) and GA<sub>3</sub> (0.5 mgL<sup>-1</sup>). Agar (0.8%) was added to medium after pH adjusted to 5.7. Medium containing BAP and IBA were autoclaved at 0.1 MPa pressure for 20 min at 121 °C, while GA<sub>3</sub> was filter-sterilized. The cultures were grown at 24±1 °C in a 16-h photoperiod at light intensity of 40 μmol m<sup>-2</sup>s<sup>-1</sup> provided by white fluorescent tubes.

Regenerated shoots (shoot/explant), regenerated shoot length (mm), internode length (mm) and hyperhydricity rate (%) were evaluated after 4 weeks of culture (Singha et al., 1987). Experimental design of this study was complete randomized design with 5 replications containing 5 explants. Data were subject to ANOVA and treatment means were compared by the Duncan's multiple range test (DMRT) at 0.05 probability level using the SAS PROC GLM (SAS Institute Inc., 1989).

## RESULTS AND DISCUSSION

The ANOVA showed that regenerated shoots and hyperhydricity rate were affected by BAP concentration, PGRs combinations and BAP concentration × PGRs combinations interaction, the regenerated shoot length by PGRs combinations and internode length by BAP concentration × PGRs combinations interaction (Table 1).

### *Proliferation*

Regenerated shoots per explants in medium supplemented with 3 mgL<sup>-1</sup> BAP was higher than medium containing 2 mgL<sup>-1</sup> BAP except BAP (2 mgL<sup>-1</sup>) + GA<sub>3</sub> (0.5 mgL<sup>-1</sup>), BAP (3 mgL<sup>-1</sup>), and BAP (2 mgL<sup>-1</sup>) + GA<sub>3</sub> (0.5 mgL<sup>-1</sup>) produced the most regenerated shoots (Table 2). It was reported that high concentrations of cytokinins led to producing more regenerated shoots in some cultivars of Asian pear (Rouzban et al., 2002). BAP (2 mgL<sup>-1</sup>) + IBA (0.1 mgL<sup>-1</sup>) were more effective than medium supplement with BAP that our results were in agreement with some workers. In *P. calleryana*, OPR 260 and OH×F 230, the highest proliferation rate was obtained in medium containing BAP (1.8 mgL<sup>-1</sup>) + IBA (0.08 mgL<sup>-1</sup>) combination (Yeo and Reed, 1995). It was reported BAP (0.5 mgL<sup>-1</sup>) + IAA (0.05 mgL<sup>-1</sup>) was more affective for proliferation in cultivars and hybrids of pear rootstocks (Nadosy, 1997). Successful proliferation reported for Japanese pear (Jinfeng and Zaosu cultivars) shoot tips with GA<sub>3</sub> pre-treatment that cultured in MS medium containing BAP (1 mgL<sup>-1</sup>) + GA<sub>3</sub> (10 mgL<sup>-1</sup>) (Zhao, 1982).

Regenerated shoots length decreased when BAP increased. More production in regenerated shoots led to less regenerated shoot elongation (Table 3). Similar results reported for Asian pears (Rouzban et al., 2002) and *P. syria* (Shibli et al., 1997) and *Pyrus* genus (Dolcet-Sanjuan, 1990). In *Pyrus*, Liaw et al. (1992) stated the highest proliferation obtained in higher BAP concentrations, while lower BAP concentrations led to

longer shoot production. The higher regenerated shoot elongation was obtained in media containing IBA (BAP + IBA and BAP + IBA + GA<sub>3</sub>) and the lowest elongation showed in BAP (Table 3). Singha (1980) reported addition of 0.1 mgL<sup>-1</sup> GA<sub>3</sub> with IAA (0.5 mgL<sup>-1</sup>) + BAP was effective for shoot elongation in *P. communis*, cv. Seckel. The effect of GA<sub>3</sub> in shoot elongation was reported on Lentil (Naeem et al., 2004).

In all combination, internode length decreased with increasing BAP concentration except for BAP + GA<sub>3</sub> that internode length increased and BAP combination this decreasing was not significant (Table 4). In *Pyrus calleryana* high concentrations of BAP led to reducing internode length of regenerated shoots (Berardi et al. 1998). The highest of internode length obtained in medium containing BAP (3 mgL<sup>-1</sup>) + GA<sub>3</sub> (0.5 mgL<sup>-1</sup>) and the lowest in medium supplement with BAP (3 mgL<sup>-1</sup>) + IBA (0.1 mgL<sup>-1</sup>) and BAP (3 mgL<sup>-1</sup>) + IBA (0.1 mgL<sup>-1</sup>) + GA<sub>3</sub> (0.5 mgL<sup>-1</sup>). The positive effect of GA<sub>3</sub> on internode elongation was reported on *Pisum sativum* (Murfet and Barber, 1961).

### *Hyperhydricity*

Hyperhydricity is a physiological disorder and malformation in cultured tissues that originated from hyper hydration and low lignifications. It is a commonly undesirable phenomenon *in vitro* culture of many plants such as pear. The Sebri hyperhydric regenerated shoots were more translucent, thick and brittle and leaves were small, compact with bright-green color. Hyperhydricity observed in all PGRs combinations but hyperhydricity rate was more affected by BAP concentration. Hyperhydricity rate increased with enhancing of BAP concentration (Table 5). It was reported hyperhydricity affected by BAP concentration, the higher concentration led to more hyperhydricity in pear (Rouzban et al., 2002; Rossi et al., 1991; Berardi et al., 1993). The lowest hyperhydricity rate (8%) obtained in BAP (2 mgL<sup>-1</sup>) + GA<sub>3</sub> (0.5 mgL<sup>-1</sup>) and it was the highest (67%) in medium containing BAP (3 mgL<sup>-1</sup>) + IBA (0.1 mgL<sup>-1</sup>) + GA<sub>3</sub> (0.5 mgL<sup>-1</sup>) (Table 5, Fig. 1). It seems in low concentration of BAP, GA<sub>3</sub> and IBA have synergic effect on hyperhydricity rate in Sebri regenerated shoots but in high concentration of BAP, different combination of PGRs did not control and reduce hyperhydricity rate.

A negative relation showed between hyperhydricity rate and internode length. In different combinations of plant growth regulators decreasing of internode length observed with increasing hyperhydricity rate (Table 5). In some Asian pear cultivars, it was observed that with increasing of BAP concentration, hyperhydricity rate increased and regenerated shoots length decreased (Rouzban et al., 2002). GA<sub>3</sub> (0.5 mgL<sup>-1</sup>) with IBA (0.1 mgL<sup>-1</sup>) induced more hyperhydricity rate rather than when one of them had been used.

Table 1. ANOVA of the influence of different combination of plant growth regulators on regenerated shoots, internode length and hyperhydricity of Sebri pear

Source	Means squares				
	Df	Regenerated shoots	Regenerated shoot length	Internode length	Hyperhydricity rate
BAp concentration	1	40.00 <sup>**</sup>	3.35 <sup>ns</sup>	1.65 <sup>ns</sup>	20930.62 <sup>**</sup>
PGRs combination	3	2.50 <sup>*</sup>	50.69 <sup>*</sup>	1.78 <sup>ns</sup>	2492.29 <sup>**</sup>
BAp concentration × PGRs combination	3	3.46 <sup>*</sup>	2.54 <sup>ns</sup>	5.74 <sup>*</sup>	1672.29 <sup>**</sup>
Error	32	0.97	18.13	1.91	117.18
Corrected total	39				

<sup>\*\*</sup>: significant at 1%, <sup>\*</sup>: significant at 5% and <sup>ns</sup>: not significant

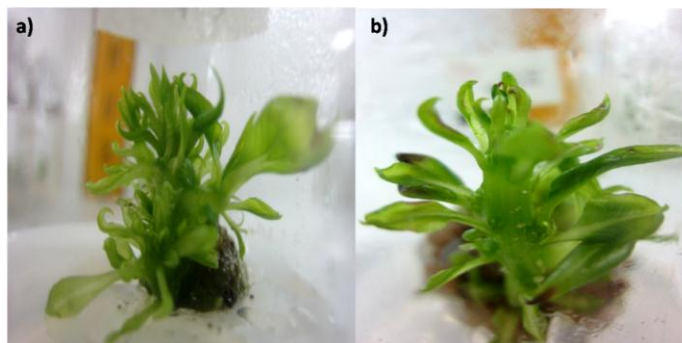


Figure 1. The effect of PGRs combination on regenerated shoot hyperhydricity a) in 2BAP + GA<sub>3</sub> and b) in 3BAP + IBA + GA<sub>3</sub>

Table 2. The effect of BAP concentration, PGRs combinations and BAP concentration × PGRs combinations interaction on regenerated shoots (shoot/explants) in Sebri pear cultivar

	BAP	PGRs combination				Average
		BAP	BAP + IBA	BAP + GA <sub>3</sub>	BAP + IBA + GA <sub>3</sub>	
BAP concentration	2	0.40 b <sup>*</sup>	1.00 b	2.60 a	0.60 b	1.15 B
	3	3.60 a	3.40 a	3.00 a	2.60 a	3.15 A
Average		2.00 AB	2.20 AB	2.80 A	1.60 B	

\* Means followed by the same letter (capital letter indicates simple and small letter indicates interaction) are not significantly different by Duncan's test at 5% probably level

Table 3. The effect of BAP concentration, PGRs combinations and BAP concentration × PGRs combinations interaction on regenerated shoot length (mm) in Sebri pear cultivar

	BAP	PGRs combination				Average
		BAP	BAP + IBA	BAP + GA <sub>3</sub>	BAP + IBA + GA <sub>3</sub>	
BAP concentration	2	3.400 a <sup>*</sup>	9.010 a	5.114 a	8.280 a	6.451 A
	3	4.224 a	8.182 a	4.382 a	6.700 a	5.872 A
Average		3.812 B	8.596 A	4.748 AB	7.490 AB	

\* Means followed by the same letter (capital letter indicates simple and small letter indicates interaction) are not significantly different by Duncan's test at 5% probably level

Table 4. The effect of BAP concentration, PGRs combinations and BAP concentration × PGRs combinations interaction on internode length (mm) in Sebri pear cultivar

	BAP	PGRs combination				Average
		BAP	BAP + IBA	BAP + GA <sub>3</sub>	BAP + IBA + GA <sub>3</sub>	
BAP concentration	2	1.427 ab <sup>*</sup>	1.733 ab	1.283 ab	2.269 ab	1.678 A
	3	1.281 ab	0.490 b	2.890 a	0.424 b	1.271 A
Average		1.354 A	1.111 A	2.086 A	1.347 A	

\* Means followed by the same letter (capital letter indicates simple and small letter indicates interaction) are not significantly different by Duncan's test at 5% probably level

Table 5. The effect of BAP concentration, PGRs combinations and BAP concentration × PGRs combinations interaction on hyperhydricity (%) in Sebri pear cultivar

	BAP	PGRs combination				Average
		BAP	BAP + IBA	BAP + GA <sub>3</sub>	BAP + IBA + GA <sub>3</sub>	
BAP concentration	2	28.00 b <sup>*</sup>	28.00 b	8.00 c	60.00 a	31.50 B
	3	63.00 a	62.00 a	59.00 a	67.00 a	62.75 A
Average		46.50 B	45.00 B	33.50 C	63.50 A	

\* Means followed by the same letter (capital letter indicates simple and small letter indicates interaction) are not significantly different by Duncan's test at 5% probably level

## CONCLUSION

The results of this experiment indicated that different combination of IBA and GA<sub>3</sub> had different results at various concentration of BAP. Higher concentrations of BAP produced higher regenerated shoots but induced higher hyperhydricity rate. Our results indicated BAP (2 mgL<sup>-1</sup>) plus GA<sub>3</sub> (0.5 mgL<sup>-1</sup>) is the most suitable PGRs combination for proliferation in Sebri pear cultivar, because of high regenerated shoot production and low hyperhydricity rate in regenerated shoots. The combination of BAP (2 mgL<sup>-1</sup>) + IBA (0.1 mgL<sup>-1</sup>) + GA<sub>3</sub> (0.5 mgL<sup>-1</sup>) induced higher hyperhydricity rate rather than other PGRs combinations. Nevertheless, hyperhydricity was affected by high concentration of BAP concentration more than PGRs combination.

## REFERENCES

- Amiri ME. 2002. Mass propagation of unique variety of pear (*Pyrus pyrifolia* (Burm) Nak. cv. Sebri by shoot tip culture *in vitro*. Acta Hort 587:555-561.
- Berardi G, Infante R, Neri D. 1998. Micropropagation of *Pyrus calleryana* Dcn. from seedlings. Scientia Hort 53: 157-165.
- Crawford RE. 1983. Nashi (Asian pear). Turners & Growers Ltd. Auckland, New Zealand
- Davarynejad GH, Davarynejad E. 2007. Comparative performance of graft incompatibility in pear / quince (*Pyrus comunis*/*Cydonia oblonga*) combination, Acta Horticulture 732: 221-227.
- Davarynejad GH, Shahriari F, Hassanpour H. 2008. Identification of graft incompatibility of pear cultivars on quince rootstock by using isozymes banding pattern and starch, Asian J Plant Sci 7 (1): 109-112.
- Dolcet-Sanjuan R, Mok DWS, Mok MC. 1990. Micropropagation of *Pyrus* and *Cydonia* and their responses to Fe-limiting conditions. Plant cell tiss org 21: 191-199.
- Dwivedi SK, Bist LD. 1999. *In Vitro* propagation of low-chill pear cv. Gola. Indian J Hort 56: 189-193.
- Erfani J, Ebadi A, Abdollahi H, Fatahi R. 2012. Genetic Diversity of Some Pear Cultivars and Genotypes Using Simple Sequence Repeat (SSR) Markers. Plant Mol Biol Rep 30: 1065-1072.
- Grattapaglia D, Machado MA. 1998. Micropropagação. In: Torres AC, Caldas LS, Buso JA, Cultura de tecidos e transformação genética de plantas. v. 1, Brasília, EMBRAPA.
- Kadota M, Niimi Y. 2003. Effects of cytokinin types and their concentrations on shoot proliferation and hyperhydricity in *in vitro* pear cultivar shoots. Plant cell tiss org 72: 261-265.
- Kadota M, Imizu K, Hirano T. 2001. Double phase *in vitro* culture using sorbitol increases shoot proliferation and reduces hyperhydricity in Japanese pear. Scientia Horticulturae 89: 207-215.
- Murashige T, Skoog F. 1962. A revised medium for rapid growth and bioassays with tobacco tissue cultures. Physiol Plant 15: 473 - 497.
- Murfet IC, Barber HN. 1961. Effects of Gibberellic Acid and Allogibberic Acid on Flowering and Internode-Length in *Pisum sativum*. Nature 191: 514 - 515.
- Naeem M, Bhatti I, Ahmad RH, Ashraf MY. 2004. Effect of some growth hormones (GA<sub>3</sub>, IAA and kinetin) on the morphology and early or delayed initiation of bud of lentil (*Lens culinaris* Medik). Pak J Bot 36(4): 801-809.
- Nadosy F. 1997. Micropropagation of pear rootstocks. Horti. Sci. 29: 17-21.
- Nosrati SZ, Zamani Z, Babalar M. 2009. Micropropagation of Four Cultivars (Dargazi, Natanzi, Shahmiveh and Williams) of Pear (*Pyrus communis* L.) . Iranian Journal of Horticultural Science 40 (2): 83-91.
- Pierik RLM. 1990. Production de plantaslibres de enfermedades In: Pierik, RLM (Ed) cultivo *in vitro* de las plantas superiores. Madrid: Ediciones Mundi-Pronsa.
- Rahemi M, Baghban R. 2002. Effects of harvesting thime and storage period on poststorage ripening of Esfahan 'Sebri' pear (*Pyrus serotina* Rehd. cv. 'Sebri'). Acta Hort 587:519-523.
- Rouzban MR, Arzani K, Moieni A. 2002. Study on *in vitro* propagation of some Asian Pear (*Pyrus serotina* Rehd.) cultivars. Seed and Plant 18:348-361.
- Safarpour M, Bahari M, Tabatabaei EBS, Abdelahi H. 2008. Determination of genetic diversity in pear (*Pyrus* spp.) using microsatellite markers. Iranian J of Horti Sci and Tech 9 (2): 113-128.
- SAS Institute Inc. 1989. SAS/STAT User's Guide, Version 6, 4th edn, SAS Institute, Inc., Cary.
- Shibli RA, Ajlouni MM, Jaradat A, Aljanabi S, Shatnawi M. 1997. Micropropagation in wild pear (*Pyrus syriaca*). Scientia Hort 68: 237-242.
- Singha S. 1980. *In vitro* propagation of 'Seckel' pear. In: Proceedings of the conference on 418 nursery production of fruit plants through tissue culture: application and feasibility: 21-22.
- Singha S, Oberly GH, Townsend FC. 1987. Changes in nutrient composition and pH of the culture medium during *in vitro* shoot proliferation of crabapple and pear. Plant cell tiss org 11: 209-220.
- Yeo DY, Reed BM. 1995. Micropropagation of *Pyrus* rootstocks. HortScience 30: 620-623.
- Zafarinia H, Arzani K, Ghasemi AA. 2010. An Evaluation of Carbohydrates and Nutrient Content of some Young Asian Pear (*Pyrus serotina* Rehd.) Cultivars on European Pear (*Pyrus communis* L.) Seedling Rootstocks under Isfahan Environmental Conditions. Iranian J of Horticultural Science 41(3): 209-221.
- Zhao HX. 1982. Shoot tip culture of pear *in vitro*. Acta Bot Sinica 24: 392-394.