Journal of Advances in Medical and Biomedical Research | ISSN:2676-6264

The Impact of Different Modes of Exercise Training on Irisin: A Systematic Review and Meta-Analysis Research

Morteza Motahari Rad¹, Nahid Bijeh^{1*}, Seyyed Reza Attarzadeh Hosseini¹, Aliakbar Raouf Saeb²

- 1. Dept. of Exercise Physiology, Faculty of Sports Sciences, Ferdowsi University of Mashhad, Mashhad, Iran
- 2. Parsian Diabetes Clinic, Mashhad, Iran

Article Info

ABSTRACT

Received: 2020/07/14; **Accepted**: 2020/09/24; **Published Online**: 30 Dec 2020;

doi) 10.30699/jambs.29.134.125

Use your device to scan and read the article online



Corresponding Information: Nahid Bijeh, Dept. of Sport Physiology, Faculty of Sports Sciences, Ferdowsi University of Mashhad, Mashhad, Iran E-Mail: : bigeh@um.ac.ir **Background & Objective:** Irisin is considered as a candidate to treat obesity and metabolic diseases. Therefore, the aim of this study was a systematic review and meta-analysis to define the effective modes of exercise on irisin concentration in adults.

Materials & Methods: Google Scholar, PubMed, Medline, Scopus, Science Direct, and references lists of articles were searched until April 25, 2020. To study the impact of exercise duration (short-term and long-term) on the pre-test and post-test concentration of irisin, 21 studies with 384 subjects were evaluated.

Results: In the present study, a significant increase was found in irisin level in long-term resistance training (LTRT) intervention groups (MD:-0.61 ng/ml, 95% CI [-1.12, 0.09], P=0.02). However, no significant changes were reported in the groups of long-term aerobic training, short-term aerobic training, and short + long term aerobic training.

Conclusion: The analysis showed that only LTRT among the types of exercise can elevate irisin concentration. Therefore, LTRT seems to have more beneficial influences, compared to other modes of exercise in inclusive irisin responses.

Keywords: Aerobic training, Irisin, Resistance training

Copyright © 2021, This is an original open-access article distributed under the terms of the Creative Commons Attribution-noncommercial 4.0 International License which permits copy and redistribution of the material just in noncommercial usages with proper citation.

Introduction

Although the beneficial effects of exercise are well documented, the molecular mechanisms by which exercise prevents chronic diseases and improves health are poorly understood. Skeletal muscles have been identified as an active endocrine organ. They have critical functions in metabolic homeostasis by releasing hormones, called myokine, during muscle contraction (1). Myokines mediate crosstalk between tissues, such as the adipose tissue, liver, skin, and pancreas. They locally affect muscles and impose metabolic impacts through autocrine, paracrine, and endocrine signaling (1).

Irisin seems to be predominantly involved in exercise adaptations as a contraction-regulated myokine. It has sufficient potential to prevent and treat obesity and metabolic diseases (2, 3). Irisin regulates energy metabolism and thermogenesis, increments white adipocytes browning (fat browning), improves insulin sensitivity, and upgrades body compositions (4). Moreover, irisin promotes mitochondrial biogenesis and effectively increases oxidative capacity (2, 3). It is produced following proteolytic cleavage and the glycosylation and dimerization of fibronectin type III domain-containing protein 5 (FNDC5), which is set via peroxisome proliferator-activated receptor (PPAR) gamma coactivator 1α (PGC- 1α) (5, 6).

It has been shown that acute or long-term exercise can induce PGC-1a expression in tissues resulting in the induction of FNDC5 production and increases irisin secretion as a myokine (7). Although various studies introduce exercise as an irisin-augmenting factor in rats (2, 8, 9), conflicting results have been reported about humans (10, 11). However, Bostrom et al. (2012) revealed irisin elevation following exercise in humans (12). In this regard, some investigations reported that short-term moderate-intensity continuous training (MICT) or high-intensity interval training (HIIT) leads to a transient increase in irisin concentration (13-15). Moreover, irisin raise has been observed in various studies in response to long-term aerobic training (AT) or resistance training (RT), while no significant impact has been reported on irisin as the result of exercise (11, 16, 17). Furthermore, it is well known that muscle contraction is essential to prevent metabolic diseases. However, the hormonal and physiological adaptations of AT and RT are different (18-20). In addition, several systematic reviews and meta-analyses have examined irisin concentration regulation by exercise. None of the previous papers included critical analyses of different modalities of exercise. In other words, it is not clear which kind of exercise is more effective in irisin concentration. Finally, this paper aimed to perform a meta-analysis to define the possible influential types of exercise training on irisin concentration in adults.

Materials and Methods

Data Sources and Searches

All English databases, including PubMed, CINAHL, Google Scholar, Medline, and Scopus were searched. The following phrases and words were searched in the keywords, titles, abstracts, introduction, subjects, and heading of articles: irisin, FNDC5, PGC1- α , exercise, physical activity, training, aerobic training, RT, concurrent training, short-term exercise training, and long-term exercise training. The Boolen search strategy was used (AND, OR, or NOT). All articles including meta-analyses, systematic reviews, and the references of the used articles were searched and assessed until April 25, 2020.

Study Selection

All types of exercise, including long-term aerobic training (LTAT), long-term resistance training (LTRT), and short-term resistance training were selected randomly in the pre- and post-test conditions. In our meta-analysis resistance exercise group entailed full-body training with a machine and weight-bearing encompassed at least six movements. Moreover, the aerobic exercise group included walking, running, and aerobic training. One of the authors evaluated the titles, treatment guidelines, summaries, and full-texts of appropriate studies.

Inclusion/Exclusion Criteria

The studies with the following criteria were selected: (1) full-text experimental and controlled trials published in English, (2) participants aged ≥ 18 years, (3) irisin was one of the main subjects of the study and was measured by enzyme-linked immunosorbent assay (ELISA), (4) the protocol of studies used AT, RT, or combined AT and RT (with an intervention period ≥ 8 weeks for long-term studies), and (5) inactive to moderately active subjects.

Studies were excluded if they met one of the following conditions: (1) review article or literature review, (2) studies with non-human participants, (3) conferences, (4) abstracts, (5) study protocols in which the participants took part in a training plan within the last six months.

Outcome Measures

The studies that had measured dynamic irisin by different ELISA kits were eligible for the present review.

Data Extraction

Titles and abstracts were assessed separately by an author (MMR). All the authors (MMR, ND, SRAH, and ARS) extracted three types of variable, including participant (study population, sample size, age, and baseline body mass index), exercise protocol details (duration, modalities, intensities, and frequency of exercise), and outcomes (irisin concentrations associated with exercise modalities). The methodology qualification of each study was evaluated separately by an author (MMR) through the Cochrane collaboration 'risk of bias' tool (21, 22).

Data Synthesis

According to all investigations, the effect size was reported for outcomes by calculating the mean differences between pre- and post-test. All outcomes were assessed independently. To compute the mean difference of irisin level, some parameters, including sample size, characteristics of the participants, methods of blood analysis, and information of exercise interventions were evaluated by the same methods in pre- and post-test conditions.

All data were assessed by Review Manager 5.3 (The Nordic Cochrane Center, Copenhagen, Denmark). The post-test mean was subtracted from the pre-test mean and the standard error of means (SEM) was changed to standard deviation. TA Tech Tip and GetData Graph Digitizer software were used for any data which had not been shown in the text or tables (displayed data in figures) and we were not able to reach the authors. Therefore, Heterogeneity was calculated as Cochrane's Q and I^2 index >50%. We presented a 5% level of significance for the forest plot to describe the results. A negative effect size demonstrated irisin concentration reduction, while a positive effect size indicated an increase. P-value<0.05 was considered statistically significant.

Study Quality

The studies were qualified by a fifteen-point tool in exercise reporting (TESTEX) scales (23). Three reviewers (MMR, ND, and SRAH) qualified the studies and reported the assessments.

Results

Overall, 384 related articles were found from Scopus, PubMed, Google Scholar, and Science Direct databases. Animal studies and pharmacologic interventions were removed leading to the final number of 214 full-text articles. Seventy studies were evaluated and 21 articles were selected to be reviewed for the moderator variables and stated inclusion criteria (PRISMA flow diagram, Figure 1). These 21 studies included 631 individuals in pre- and post-test. Moreover, 457, 298, 159, and 174 participants were in the aerobic training (short-term + long-term aerobic training) (SLTAT), short-term aerobic training (STAT), LTAT, and LTRT groups, respectively.

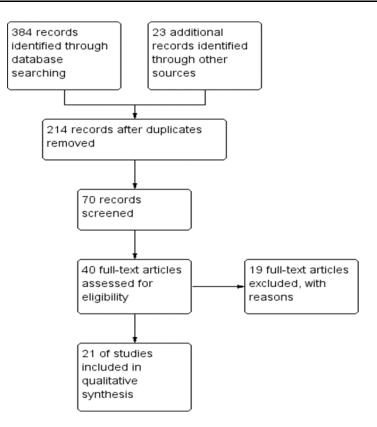
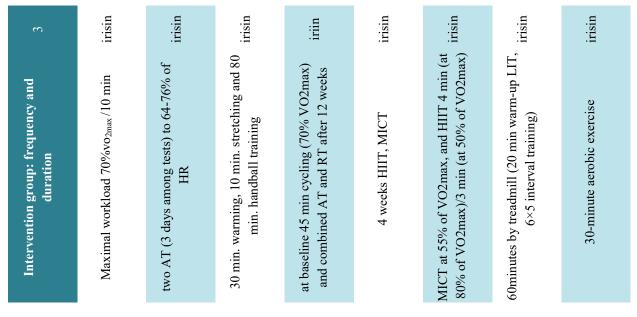


Figure 1. The PRISMA flow chart

Intervention Details

Studies duration range was 24 hours to 12 weeks. Full article characteristics have been shown in <u>Table 1</u>. The aerobic exercise training interventions had a range of 30-60 min with an intensity of 50%-85% VO_{2max} . In the resistance exercise intervention, it ranged from 40%-90% 1RM.

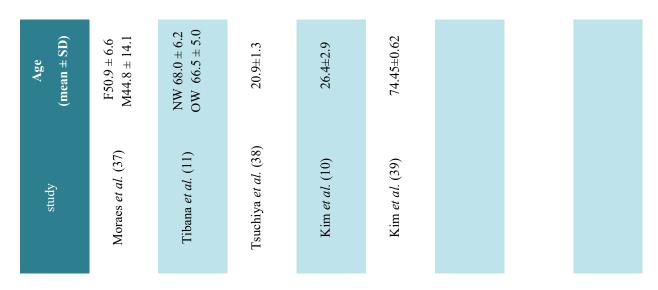
Table 1. The characteristics of the investigated articles



Assessment measure/units	study	Age (mean±SD)	BMI (mean ± SD)	Disease	gender	N EX	Modes of exercise
irisin	Daskalopoulou <i>et al.</i> (24)	23.0±3.3	22.4±2.5	None	F 15 M 20	35	Short-term aerobic
irisin	Algul <i>et al.</i> (25)	19.2±0.7	21.3±0.4	None	X	30	short-term aerobic
irisin	Dundar et aql. (26)	16.052±0.848	23.089±1.090	None	X	19	long-term
iri	Norheim et al. (27)	40-65	NW 23.5±2.0 OW 29.0±2.4	None Overweight	М	26	long-term aerobic
irisin	Dunnwald <i>et al.</i> (28)	59.6±5.7	27.8 ± 2.8	T2DM	4 F 10 M	14	long-term aerobic
irisin	Winn <i>et al.</i> (29)	24.3±1.4	37.3±2.1	opese	ц	Ξ	short-term aerobic
irisin	Nygard <i>et al.</i> (15)	32±9	24.5±2.4	None	2 F 7M	6	Short-term aerobic
irisin	Anastasilakis <i>et al.</i> (30)	20.0 0.1	MM	None	F 61 M 61	122	Short-term aerobic

Age (mean ± SD)	BMI (mean ± SD)	Disease	gender	N EX	Modes of exercise	Intervention group: frequency and duration
23 ± 3.3	22 ± 5.6	None	ц	38	Short-term aerobic	Bruce's protocol was conducted from Monday to Friday during 2 weeks
27.4±3.8	22.0±1.1	None	F9 M8	17	short-term aerobic	an acute exercise test at an intensity corresponding to 80% of their VO2max for 50 min.
F 3.8 ± 4.7 M 2.71 ± 1.6	F 5.2 ± 4.97 M 4.29 ± 2.94	None	F 5 M 7	12	Short-term aerobic	90 min treadmill training with 54 min at 60 % of VO 2 max
HIIT 16.13±1.64 A 15.47±1.73	HIIT 31.65±4.83 A 32.67±5.91	Obese	ц	30	Long-term aerobic	Cycling at 65% of HR for 40 min. 1 min training and 1 min rest classified by 6 rounds at 85–95% HRpeak.
38.4±8.2	HFL 30.1±4.0 LFL 35.7±4.5	Obese	Μ	12	Short-term aerobic	10 session in cryogenic chamber
62.3 ± 3.5	26.18±5.5	None	М	10	Long-term resistance	RT protocol included lower muscles training and core muscles
29.37±5.14	27.13±2.16	None	Μ	r	Long-term resistance	26 exercises were divided into 5 sets with 10-15 movements at 60-80% 1RM for 8 weeks.
LRT 25.27±3.34 HRT 23.50 LRT 23.8±1.58 HRT ± 2.27 ± 2.37 23.6±1.57	LRT 23.8±1.58 HRT 23.6±1.57	None	ц	21	Long-term resistance	LRT (40%-60%1RM and 20-30 repetition) HRT (70%-90% 1RM and 5-15 repetition)

BMI (mean ± SD)	Disease	gender	EX	Modes of exercise	Intervention group: frequency and duration	Assessment measure/units	study
F 24.2 ± 2.7 M23.5 ± 3.9	hemodialys 11 female is male	l female 7 male	18	Long-term resistance	72sessions, lower limbs training with 60%-70%1RM	irisin	Saldaña <i>et al.</i> (31)
NW 24.3±3.6 OW 30.9±3.1	MO MN	female	49	Long-term resistance	16 weeks, 10 exercises, 3 sets, 6-12 repetitions maximum, 30-s rest between sets	irisin	Qiu <i>et al.</i> (13)
21.0±2.9	None	male	20	Long-term resistance	4weeks, 3 consecutive 30-s maximal pedaling exercise with 10-min rest between sets	İrisin	Kraemer <i>et al.</i> (32)
27.0±3.4	MO	3female 7male	10	Long-term resistance	8 weeks, 60-min 6 movements, 65-80 % 1RM, 10-12 repetitions for 3 sets (2-3 s concentric, 2-3 s eccentric)	iriin	Herrera <i>et al.</i> (33)
25.28	None	female	18	long-term resistance	12 weeks(warm-up, strength elasticity training for 40 min cooldown)	irisin	Dulian <i>et al.</i> (34)
							Zhao <i>et al.</i> (35)
							Bang <i>et al.</i> (16)
							Moienneia <i>et</i> al. (36)



Outcomes Measures

Change in Irisin

Irisin changes have been reported by 21 studies, which included a total of 631 members (26 pre- and

post-tests). We mixed the outcomes to make the random-effects model. No considerable difference in irisin level was observed in the post-test training (MD:-0.36 ng/mL, 95%CI [-0.80, 0.07], P=0.1) (Figure 2).

	pr	e-test		po	st-test			Std. Mean Difference		Std. Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Random, 95% Cl	Year	IV, Random, 95% Cl
Nurheim N 2013	125	10	26	110	15	26	4.2%	1.16 [0.57, 1.75]	2013	
Norheim OW 2013	190	20	26	180	20	26	4.2%	0.49 [-0.06, 1.04]	2013	—
Kraemer 2013	62.08	4.1	12	74.1	2.3	12	3.2%	-3.49 [-4.84, -2.15]	2013	←
Moraes 2013	23.5	3.9	18	24	4.3	18	4.1%	-0.12 [-0.77, 0.53]	2013	
Anastasilakis 2014	74.8	6.6	122	78.1	7.4	122	4.5%	-0.47 [-0.72, -0.21]	2014	
Bang 2014	1,461.7	219.88	7	1,384	159.49	7	3.6%	0.38 [-0.68, 1.44]	2014	
Dulian HFL 2015	105.3	5.3	12	94.9	1.2	12	3.5%	2.61 [1.48, 3.75]	2015	
Kim 2015	726	50.3	10	850	60.8	10	3.5%	-2.13 [-3.27, -0.98]	2015	
Kim2015	65	20	18	85	20	18	4.1%	-0.98 [-1.67, -0.28]	2015	
Dulian LFL 2015	117.7	37.5	12	104.8	21.7	12	3.9%	0.41 [-0.40, 1.22]	2015	
Nygard 2015	428	66	9	437	56	9	3.8%	-0.14 [-1.07, 0.79]	2015	
Moienneia HRT 2016	78.1	39.7	21	68.8	31.1	21	4.2%	0.26 [-0.35, 0.86]	2016	_
Moienneia LRT 2016	86.9	39	21	71	47.2	21	4.2%	0.36 [-0.25, 0.97]	2016	
Tibana 2016	184.1	72.5	49	228.2	59.5	49	4.4%	-0.66 [-1.07, -0.25]	2016	_ —
Tsuchiya 2016	5.3	1.3	20	6.8	1	20	4.1%	-1.27 [-1.95, -0.58]	2016	
Algul 2017	3.858	0.11	30	4.056	1.12	30	4.3%	-0.25 [-0.75, 0.26]	2017	
Herrera HIIT 2017	3.59	1.29	30	3.7	1.26	30	4.3%	-0.09 [-0.59, 0.42]	2017	
Herrera A 2017	3.66	0.8	30	3.56	0.69	30	4.3%	0.13 [-0.37, 0.64]	2017	
zhao 2017	287	143.5	10	556	126.6	10	3.5%	-1.90 [-3.00, -0.81]	2017	
winn 2017	1.02	0.3	11	1.04	0.4	11	3.9%	-0.05 [-0.89, 0.78]	2017	
Daskalopoulou 2018	84.8	36.1	35	113.3	52.7	35	4.3%	-0.62 [-1.10, -0.14]	2018	
Qiu A 2018	71.98	13.69	17	79	1.075	17	4.1%	-0.71 [-1.40, -0.01]	2018	
Dundar 2019	698.334	237.7	19	1,344.495	385.043	19	4.0%	-1.98 [-2.77, -1.19]	2019	
Dunwald MIT 2019	0.57	0.04	14	0.61	0.06	14	4.0%	-0.76 [-1.53, 0.01]	2019	
Dunwald HIIT2019	0.6	0.06	14	0.58	0.07	14	4.0%	0.30 [-0.45, 1.04]	2019	
Saldana 2019	158	0.1	38	211	0.1	38	0.0%	-524.61 [-610.26, -438.96]	2019	•
Total (95% CI)			631			631	100.0%	-0.36 [-0.80, 0.07]		•
Heterogeneity: Tau ² = 1	.09; Chi ² =	296.81,	df = 25	(P < 0.00001	l); l² = 929	6				
Test for overall effect: Z										-Z -1 0 1 2
		-,								Favours [experimental] Favours [control]

Figure 2. The forest plot for irisin level in exercise intervention

Exercise Modality Investigation

We assessed the effects of intervention training period (i.e., LTAT, STAT, and LTRT) on irisin level. This analysis found that irisin level had not been changed by SLTAT significantly (MD:-0.21 ng/ml, 955 CI [-0.83, 0.4], P=0.49). The obtained values were

as LTAT (MD:-0.08 ng/mL, 95% CI [-0.72, 0.57], P=0.82) and STAT (MD:-0.34 ng/mL, 95% CI [-1.35, 0.67], P=0.51). However, the values may alter in LTRT significantly (MD:-0.61 ng/mL, 95% CI [-1.12, 0.09], P=0.02). In contrast, the irisin level increased after LTRT (Figures 3-6).

Mean 1.02 158	SD 0.3	Total	Mean	SD	Tetel			
	0.3			50	Total	Weight	IV, Random, 95% Cl	IV, Random, 95% Cl
158		11	1.04	0.4	11	6.1%	-0.05 [-0.89, 0.78]	
130	0.1	38	211	0.1	38	0.0%	-524.61 [-610.26, -438.96]	•
71.98	13.69	17	79	10.75	17	6.3%	-0.56 [-1.24, 0.13]	
428	66	9	437	56	9	5.9%	-0.14 [-1.07, 0.79]	
125	10	26	110	15	26	6.5%	1.16 [0.57, 1.75]	
190	20	26	180	20	26	6.5%	0.49 [-0.06, 1.04]	
62.08	4.1	12	74.1	2.3	12	5.2%	-3.49 [-4.84, -2.15]	←
3.59	1.29	30	3.7	1.26	30	6.6%	-0.09 [-0.59, 0.42]	
3.66	0.8	30	3.56	0.69	30	6.6%	0.13 [-0.37, 0.64]	_ -
0.6	0.06	14	0.58	0.07	14	6.2%	0.30 [-0.45, 1.04]	_ +-
0.57	0.04	14	0.61	0.06	14	6.2%	-0.76 [-1.53, 0.01]	
698.334	237.7	19	1,344.495	385.043	19	6.2%	-1.98 [-2.77, -1.19]	
117.7	37.5	12	104.8	21.7	12	6.1%	0.41 [-0.40, 1.22]	_ + •
105.3	5.3	12	94.9	1.2	12	5.6%	2.61 [1.48, 3.75]	
84.8	36.1	35	113.3	52.7	35	6.6%	-0.62 [-1.10, -0.14]	
74.8	6.6	122	78.1	7.4	122	6.8%	-0.47 [-0.72, -0.21]	-
3.858	0.11	30	4.056	1.12	30	6.6%	-0.25 [-0.75, 0.26]	
		457			457	100.0%	-0.21 [-0.83, 0.40]	-
2: Chi ² =	252.52	df = 16	6 (P < 0.000)	01); I ² = 94	%			<u> </u>
								-2 -1 0 1 2 pre-test post-test
	428 125 190 62.08 3.66 0.67 698.334 117.7 105.3 84.8 74.8 3.858 2; Chi ² =	428 66 125 10 190 20 62.08 4.1 3.59 1.29 3.66 0.8 0.6 0.06 0.57 0.04 698.334 237.7 117.7 37.5 105.3 5.3 84.8 36.1 74.8 6.6 3.858 0.11	428 66 9 125 10 26 190 20 26 62.08 4.1 12 3.59 1.29 30 3.66 0.8 30 0.6 0.06 14 057 0.04 14 698.334 237.7 19 117.7 37.5 12 105.3 5.3 12 84.8 36.1 35 74.8 6.6 122 3.858 0.11 30 457 2; Chi² = 252.52, df = 16	428 66 9 437 125 10 26 110 190 20 26 180 62.08 4.1 12 74.1 3.59 1.29 30 3.7 3.66 0.8 30 3.56 0.57 0.04 14 0.61 698.334 237.7 19 1,344.495 117.7 37.5 12 104.8 105.3 5.3 12 94.9 84.8 36.1 35 113.3 74.8 6.6 122 78.1 3.858 0.11 30 4.056 457 2; ChI ² = 252.52, df = 16 (P < 0.000	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$

Figure 3. The forest plot of irisin level in an aerobic training group

	pr	e-test		ро	st-tes	t	:	Std. Mean Difference		Std. Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Random, 95% Cl	Year	IV, Random, 95% Cl
Nurheim N 2013	125	10	26	110	15	26	14.6%	1.16 [0.57, 1.75]	2013	_ _
Norheim OW 2013	190	20	26	180	20	26	14.8%	0.49 [-0.06, 1.04]	2013	—
Herrera A 2017	3.66	0.8	30	3.56	0.69	30	15.1%	0.13 [-0.37, 0.64]	2017	
Herrera HIIT 2017	3.59	1.29	30	3.7	1.26	30	15.1%	-0.09 [-0.59, 0.42]	2017	_ - -
Dunwald HIIT2019	0.57	0.04	14	0.61	0.06	14	13.5%	-0.76 [-1.53, 0.01]	2019	
Dundar 2019	698.33	237	19	1,344	385	19	13.3%	-1.98 [-2.77, -1.19]	2019	_
Dunwald MIT 2019	0.6	0.06	14	0.58	0.07	14	13.6%	0.30 [-0.45, 1.04]	2019	- -
Total (95% CI)			159			159	100.0%	-0.08 [-0.72, 0.57]		-
Heterogeneity: Tau ² =	= 0.66; Ch	i² = 46	.30, df=	= 6 (P <	0.000	01); I ^z =	87%			
Test for overall effect	-									-2 -1 U 1 2 pre-test post-test

Figure 4. The forest plot of irisin level in LTAT group

	р	re-test		p	ost-test			Std. Mean Difference	Std. Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Random, 95% Cl	IV, Random, 95% Cl
winn 2017	1.02	0.3	11	1.04	0.4	11	11.1%	-0.05 [-0.89, 0.78]	_
Saldana 2019	158	0.1	38	211	0.1	38	0.0%	-524.61 [-610.26, -438.96]	•
Qiu A 2018	71.98	13.69	17	79	10.75	17	11.4%	-0.56 [-1.24, 0.13]	
Nygard 2015	428	66	9	437	56	9	10.9%	-0.14 [-1.07, 0.79]	
Kraemer 2013	62.08	4.1	12	74.1	2.3	12	9.9%	-3.49 [-4.84, -2.15]	←
Dulian LFL 2015	117.7	37.5	12	104.8	21.7	12	11.1%	0.41 [-0.40, 1.22]	
Dulian HFL 2015	105.3	5.3	12	94.9	1.2	12	10.4%	2.61 [1.48, 3.75]	
Daskalopoulou 2018	84.8	36.1	35	113.3	52.7	35	11.7%	-0.62 [-1.10, -0.14]	
Anastasilakis 2014	74.8	6.6	122	78.1	7.4	122	11.9%	-0.47 [-0.72, -0.21]	-
Algul 2017	3.858	0.11	30	4.056	1.12	30	11.6%	-0.25 [-0.75, 0.26]	-+
Total (95% CI)			298			298	100.0%	-0.34 [-1.35, 0.67]	
Heterogeneity: Tau² = 2				:9 (P <	0.00001); I² = 9	15%		
Test for overall effect: Z	(= 0.66	P = 0.51	1)						pre-test post-test

Figure 5. The forest plot of irisin level in STAT group

	p	re-test		р	ost-test			Std. Mean Difference		Std. Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Random, 95% Cl	Year	IV, Random, 95% Cl
Moraes 2013	23.5	3.9	18	24	4.3	18	12.0%	-0.12 [-0.77, 0.53]	2013	
Bang 2014	1,461.7	219.88	7	1,384	159.49	7	9.1%	0.38 [-0.68, 1.44]	2014	
Kim2015	726	50.2	10	850	60.8	10	8.6%	-2.13 [-3.28, -0.99]	2015	
Kim 2015	65	20	18	85	20	18	11.7%	-0.98 [-1.67, -0.28]	2015	_ -
Moienneia HRT 2016	78.1	39.7	21	68.8	31.1	21	12.3%	0.26 [-0.35, 0.86]	2016	- -
Moienneia LRT 2016	86.9	39	21	71	47.2	21	12.3%	0.36 [-0.25, 0.97]	2016	+
Tibana 2016	184.1	72.5	49	228.2	59.5	49	13.6%	-0.66 [-1.07, -0.25]	2016	
Tsuchiya 2016	5.3	1.3	20	6.8	1	20	11.7%	-1.27 [-1.95, -0.58]	2016	_ -
zhao 2017	287	143.5	10	556	126.6	10	8.9%	-1.90 [-3.00, -0.81]	2017	
Total (95% CI)			174			174	100.0%	-0.61 [-1.12, -0.09]		•
Heterogeneity: Tau ² = 0	.47; Chi²=	: 38.83, d	if = 8 (F	< 0.000	001); I ² =	79%				
Test for overall effect: Z	= 2.31 (P	= 0.02)								-2 -1 U 1 2 pre-test post-test
										pre-rest post-rest

Figure 6. The forest plot of irisin level in LTRT intervention

Variables	Type of exercise				All
	Aerobic	Long-term A	Short-term A	Long-term R	
Irisin					
N. studies	13	4	9	8	21
Mean difference(MD)	-0.21	-0.08	-0.34	-0.61	-0.36
95% CI	[-0.83, 0.40]	[-0.72, 0.57]	[-1.35, 0.67]	[-1.12, 0.09]	[-0.80, 0.07]
p-value	0.49	0.82	0.51	0.02	0.10
I ² (%)	94	87	95	79	92

Table 2. subgroup searching result in meta-analysis

Heterogeneity and Publication Bias

The data in all groups revealed high heterogeneity ($I^2=92\%$, P=0.1), and the heterogeneity of groups are shown below. The publication risk of bias was determined by Egger plot and it was low (<u>Figure 4</u>).

Study Quality

The quality of all investigated articles was reported by the TESTEX with the results of moderate to good with a median score of 10.14 (range: 7-13) out of the maximum score of 15 (<u>Table 3</u>). All 7, 13, 7, and 5 studies met the criteria of physical activity monitoring, intention to treat analyses, similarities training intensity, assessor blinding criterion, and allocation concealment. In addition, half of the trials met other TESTEX criteria.

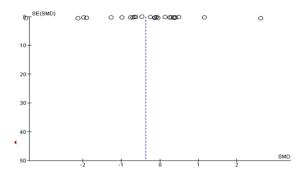


Figure 7. Funnel plot of aerobic and resistance exercises

Overall ESTEX (15)	12	10	10	7	11	10	12	8	12	10	10	7	6	10	12	10	10	10	8	13	12
úxercise volume & Energy expenditure	0	1	1	1	1	-	1	1	-	1	0	0	1	1	1	0	0	1	0	1	1
Relative exercise intensity constant	-		-	1	1	-	0	0	1	0	0	1	1	-	1	0	0	0	1	0	-
Activity Ionitoring in Introl group	0	0	0	0	0	0	1	1	0	0	0	0	0	1	0	1	0	1	1	1	0
oint measures & measures of variability	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
Reporting between group statistical	7	2	2	2	7	2	2	2	7	2	2	2	2	2	7	2	7	7	2	2	2
ntention to treat analysis	-	0	0	0	0	0	0	0	1	0	1	0	0	0	1	1	0	0	0	1	-

Table 3. Table 3. This table has been developed to evaluate the quality of articles in the field of exercise studies (TESTEX)

Study	Eligibility Criteria specified	andomizati _A llocation on details oncealed specified	d Allocation oncealed	Groups similar at baseline	Assessors blinded	Outcomes measures ssessed > 85% participants#
)askalopoulou <i>et al</i> .	1	1	0	1	1	3
Algul <i>et al.</i>	1	1	0	1	0	2
Dundar <i>et al</i> .	1	1	0	1	0	2
Norheim et al.	1	0	0	0	0	1
Dunwald <i>et al</i> .	1	1	0	1	0	3
Winn <i>et al</i> .	1	1	0	1	0	2
Nygard <i>et al</i> .	1	1	1	1	1	2
Anastasilakis <i>et al</i> .	1	0	0	1	0	1
Saldana <i>et al</i> .	1	0	0	1	-	Э
Qiu et al.	1	1	-	1	0	2
Kraemer et al.	1	1	0	1	0	3
Herrera <i>et al.</i>	1	0	0	0	0	2
Dulian <i>et al</i> .	1	1	0	1	0	1
Zhao <i>et al</i> .	-	0	0	-1	0	2
Bang <i>et al</i> .	1	1	-	1	0	2
Moienneiaa et al.	1	1	0	0	1	2
Moaress et al.	1	1	1	1	1	7
Tibana <i>et al.</i>	1	0	0	1	0	3
Tsuchiya <i>et al</i> .	1	0	0	1	0	1
Kim et al.	-			-1	1	2
Kim et al.	1	1	0	1	1	2

Discussion

This study aimed to investigate the influences of several exercise modalities on irisin concentration in adults. The results showed that LSTAT, LTAT, and STAT did not have any impact on irisin, while LTRT increased irisin concentration in adults. Although irisin is known as an exercise-induced myokine, inconsistent results have been reported in humans (11, 16, 17).

A meta-analysis indicated that chronic exercise training leads to a significant reduction in irisin concentration in the randomized controlled trials, while evidence remains inconclusive in the non-randomized studies (22). Another meta-analysis reported that irisin concentration increased immediately after an acute session of exercise in adults (40). In these two mentioned papers, the irisin level alterations were studied following three (AT + RT + concurrent training) or two types (AT + RT) of exercise.

In the present study, the effect of AT and RT on irisin concentration was examined separately. In line with our results, a study observed that acute or chronic endurance exercise did not affect irisin concentration (41). It has been recognized that AT induced PGC-1 α , which has an important role in irisin production. Some evidence has pointed to independent irisin raise and these studies require to be more explored and validated (8).

Moreover, lean mass, fat mass, and glucose concentration have been observed to be associated with irisin levels (42). It has been shown that irisin concentration is positively correlated with an enhanced risk of insulin resistance and metabolic diseases (5). Therefore, it has been reported that irisin is not only

released by muscles but also by adipose tissue. It seems that adipose tissue reduction by AT could affect irisin concentration (43). In other words, a reduction in adipose tissue as an irisin target tissue and the origin of irisin secretion can modulate its concentration (7). It is a candidate to treat obesity and type 2 diabetes and can elevate insulin sensitivity by diminishing endoplasmic reticulum stress and improving the survival of pancreatic beta cells and GLUT4 (42).

Irisin promotes fat browning by upregulating the uncoupling protein 1 (UCP1) expression (4). It can stimulate mitochondrial biogenesis by improving the expression of PPARA and mitochondrial transcription factor A (TFAM) (2, 3). It has been reported that irisin can modulate the function of diverse organs, such as cardiac and skeletal muscles, liver, bone, brain, adipose tissue, pancreas, ovary, and kidney at different levels (7).

Irisin not only enhances the oxidative capacity of muscle but also can stimulate muscle hypertrophy by affecting the Akt/mTOR/p70S6 pathway (44). Although it is released from various tissues, the skeletal muscles can secrete more than 70% of irisin in humans (45). Consequently, an increase in muscle mass by RT seems to affect irisin concentration (8). Consistent with our findings, a study observed that eight-week RT can elevate irisin level, while AT did not change its concentration significantly. RT is suggested as an efficient exercise type in overweight/obese people considering the increased irisin concentration (10).

Zhao et al. (2017) found serum irisin increment after 12-weeks of RT (35). Meantime, cross-sectional investigations have determined that irisin concentration is positively associated with insulin-like growth factor-1 (IGF-1), and the strength and circumference of biceps (44, 46, 47). In comparison, the irisin level is negatively correlated with age, cholesterol, insulin, and adiponectin concentration. Irisin potential has been shown in metabolic homeostasis (9, 44). Similarly, irisin raise has been reported through IGF-1 secretion during RT which promotes cell proliferation and myogenesis (44, 46, **48)**.

With all this in mind, it has been reported that skeletal muscle development by RT can increase myokines release (49, 50). Although the upregulation of PGC1 α is an essential factor for irisin release, intracellular muscle ATP deprivation by muscle contraction and SMAD2/3 suppression by follistatin seem to stimulate irisin secretion following RT (2, 3). However, there is no sufficient knowledge to elucidate the mechanism of exercise-induced irisin.

Furthermore, the present study had some limitations. Firstly, several studies have been performed on animals and there is little knowledge about humans. Second, our ability was limited due to the low number of investigations which have studied acute or chronic AT and RT effects on irisin concentration in human. Third, some studies have used several types of medications but our primary search was limited to English ones. Therefore, we were not able to extract all data and to obtain all potentially relevant studies.

Conclusion

The current review and meta-analysis showed that SLTAT, STAT, and LTAT cannot affect irisin concentration. However, it was found that LTRT can augment irisin concentration.

Acknowledgments

This study is based on research project No. 46208, which was sponsored by Ferdowsi University of Mashhad, Iran.

Conflict of Interest

The authors notified that there are no conflicts of interest about the publication of this article.

References

- Pedersen BK, Febbraio MA. Muscles, exercise and obesity: skeletal muscle as a secretory organ. Nature Rev Endocrinol. 2012;8(8):457-65. [DOI:10.1038/nrendo.2012.49]
- Yang D, Yang Y, Li Y, Han R. Physical exercise as therapy for type 2 diabetes mellitus: From mechanism to orientation. Ann Nutr Metab. 2019;74(4):313-21. [DOI:10.1159/000500110]
- Lee P, Linderman Joyce D, Smith S, et al. Irisin and FGF21 are cold-induced endocrine activators of brown fat function in humans. Cell Metab. 2014;19(2):302-9.
 [DOI:10.1016/j.cmet.2013.12.017]
- Son JS, Chae SA, Testroet ED, Du M, Jun H-P. Exercise-induced myokines: a brief review of controversial issues of this decade. Expert Rev Endocrinol Metab. 2018;13(1):51-8.
 [DOI:10.1080/17446651.2018.1416290]
- Choi YK, Kim MK, Bae KH, et al. Serum irisin levels in new-onset type 2 diabetes. Diabet Res Clin Pract. 2013;100(1):96-101.
 [DOI:10.1016/j.diabres.2013.01.007]
- Kurdiova T, Balaz M, Vician M, et al. Effects of obesity, diabetes and exercise on Fndc5 gene expression and irisin release in human skeletal muscle and adipose tissue: in vivo and in vitro studies. J Physiol. 2014;592(5):1091-107. [DOI:10.1113/jphysiol.2013.264655]
- Chen N, Li Q, Liu J, Jia S. Irisin, an exerciseinduced myokine as a metabolic regulator: an updated narrative review. Diabet Metab Res Rev. 2016;32(1):51-9. [DOI:10.1002/dmrr.2660]
- Behera J, Ison J, Tyagi N. Exercise improved cognitive impairment in aging mice via skeletal irisin. FASEB J. 2020;34(S1):1 [DOI:10.1096/fasebj.2020.34.s1.05839]
- Laurens C, Bergouignan A, Moro C. Exercisereleased myokines in the control of energy metabolism. Front Physiol. 2020;11:91. [DOI:10.3389/fphys.2020.00091]
- Kim HJ, Lee HJ, So B, Son JS, Yoon D, Song W. Effect of aerobic training and resistance training on circulating irisin level and their association with change of body composition in overweight/obese adults: a pilot study. Physiol Res. 2016;65(2):271. [DOI:10.33549/physiolres.932997]
- Tibana RA, da Cunha Nascimento D, Frade de Souza NM, et al. Irisin levels are not associated to resistance training-induced alterations in body mass composition in older untrained women with and without obesity. J Nutr Health Age. 2017;21(3):241-6. [DOI:10.1007/s12603-016-0748-4]

- 12. Boström P, Wu J, Jedrychowski MP, et al. A PGC1-[agr]-dependent myokine that drives brown-fat-like development of white fat and thermogenesis. Nature. 2012;481(7382):463-8. [DOI:10.1038/nature10777]
- Qiu S, Bosnyák E, Treff G, et al. Acute exerciseinduced irisin release in healthy adults: Associations with training status and exercise mode. Europ J Sport Sci. 2018;18(9):1226-33. [DOI:10.1080/17461391.2018.1478452]
- Eaton M, Granata C, Barry J, Safdar A, Bishop D, Little JP. Impact of a single bout of high-intensity interval exercise and short-term interval training on interleukin-6, FNDC5, and METRNL mRNA expression in human skeletal muscle. J Sport Health Sci. 2017;7:1-6. [DOI:10.1016/j.jshs.2017.01.003]
- Nygaard H, Slettaløkken G, Vegge G, et al. Irisin in blood increases transiently after single sessions of intense endurance exercise and heavy strength training. PLOS ONE. 2015;10(3):e0121367. [DOI:10.1371/journal.pone.0121367]
- Bang HS, Seo DY, Chung YM, et al. Ursolic acidinduced elevation of serum irisin augments muscle strength during resistance training in men. Korean J Physiol Pharmacol. 2014;18(5):441-6.
 [DOI:10.4196/kjpp.2014.18.5.441]
- Planella-Farrugia C, Comas F, Sabater-Masdeu M, et al. Circulating irisin and myostatin as markers of muscle strength and physical condition in elderly subjects. Front Physiol. 2019;10(871). [DOI:10.3389/fphys.2019.00871]
- Atherton PJ, Babraj J, Smith K, Singh J, Rennie MJ, Wackerhage H. Selective activation of AMPK-PGC-1α or PKB-TSC2-mTOR signaling can explain specific adaptive responses to endurance or resistance training-like electrical muscle stimulation. FASEB J. 2005;19(7):786-8.
 [DOI:10.1096/fj.04-2179fje]
- Eckardt K, Gorgens SW, Raschke S, Eckel J. Myokines in insulin resistance and type 2 diabetes. Diabetologia. 2014;57(6):1087-99.
 [DOI:10.1007/s00125-014-3224-x]
- Rodríguez A, Becerril S, Ezquerro S, Méndez-Giménez L, Frühbeck G. Crosstalk between adipokines and myokines in fat browning. Acta Physiologica. 2017;219(2):362-81. [DOI:10.1111/apha.12686]
- Mohammad Rahimi GR, Smart NA, Liang MTC, et al. The impact of different modes of exercise training on bone mineral density in older postmenopausal women: A systematic review and meta-analysis research. Calcified Tissue Int. 2020;106(6):577-90. [DOI:10.1007/s00223-020-00671-w]

- Qiu S, Cai X, Sun Z, Schumann U, Zügel M, Steinacker JM. Chronic exercise training and circulating irisin in adults: A meta-analysis. Sports Med. 2015;45(11):1577-88. [DOI:10.1007/s40279-014-0293-4]
- 23. Smart N, Waldron M, Ismail H, et al. Validation of a new tool for the assessment of study quality and reporting in exercise training studies: TESTEX. Int J Evid-Based Health Care. 2015;13:9-18.
 [DOI:10.1097/XEB.000000000000020]
- Daskalopoulou S, Alexandra BC, Yessica-Haydee G, et al. Plasma irisin levels progressively increase in response to increasing exercise workloads in young, healthy, active subjects. Europ J Endocrinol. 2014;171(3):343-52. [DOI:10.1530/EJE-14-0204]
- Algul S, Ozdenk C, Ozcelik O. Variations in leptin, nesfatin-1 and irisin levels induced by aerobic exercise in young trained and untrained male subjects. Biol Sport. 2017;34(4):339-44.
 [DOI:10.5114/biolsport.2017.69821]
- 26. Dündar A. Effect of Acute Handball Training on Irisin, Leptin and Some Biochemical Parameters for Adolescence Handball Players. Univers J Educa Res. 2019;7(2):318-22. [DOI:10.13189/ujer.2019.070202]
- Norheim F, Langleite TM, Hjorth M, et al. The effects of acute and chronic exercise on PGC-1α, irisin and browning of subcutaneous adipose tissue in humans. FEBS J. 2014;281(3):739-49.
 [DOI:10.1111/febs.12619]
- Dünnwald T, Melmer A, Gatterer H, et al. Supervised short-term high-intensity training on plasma irisin concentrations in type 2 diabetic patients. Int J Sport Med. 2019;40(03):158-64.
 [DOI:10.1055/a-0828-8047]
- Winn NC, Grunewald ZI, Liu Y, Heden TD, Nyhoff LM, Kanaley JA. Plasma irisin modestly increases during moderate and high-intensity afternoon exercise in obese females. PloS one. 2017;12(1):1-12. [DOI:10.1371/journal.pone.0170690]
- Anastasilakis AD, Polyzos SA, Saridakis ZG, et al. Circulating irisin in healthy, young individuals: day-night rhythm, effects of food intake and exercise, and associations with gender, physical activity, diet, and body composition. J Clin Endocrinol Metab. 2014;99(9):3247-55.
 [DOI:10.1210/jc.2014-1367]
- 31. de la Torre-Saldaña VA, Gómez-Sámano MÁ, Gómez-Pérez FJ, et al. Fasting insulin and alanine amino transferase, but not FGF21, were independent parameters related with irisin increment after intensive aerobic exercising. Revista de Investigación Clínica.

2019;71(2):133-40. [DOI:10.24875/RIC.18002764]

- 32. Kraemer RR, Shockett P, Webb ND, Shah U, Castracane VD. A transient elevated irisin blood concentration in response to prolonged, moderate aerobic exercise in young men and women. Horm Metab Res. 2014;46(02):150-4. [DOI:10.1249/01.mss.0000494385.02684.8b]
- Archundia-Herrera C, Macias-Cervantes M, Ruiz-Muñoz B, Vargas-Ortiz K, Kornhauser C, Perez-Vazquez V. Muscle irisin response to aerobic vs HIIT in overweight female adolescents. Diabet Metab Synd. 2017;9(1):101. [DOI:10.1186/s13098-017-0302-5]
- Dulian K, Laskowski R, Grzywacz T, et al. The whole body cryostimulation modifies irisin concentration and reduces inflammation in middle aged, obese men. Cryobiology. 2015;71(3):398-404.
 [DOI:10.1016/j.cryobiol.2015.10.143]
- Zhao J, Su Z, Qu C, Dong Y. Effects of 12 weeks resistance training on serum irisin in older male adults. Front Physiol. 2017;8:171.
 [DOI:10.3389/fphys.2017.00171]
- Moienneia N, Attarzadeh Hosseini SR. Acute and chronic responses of metabolic myokine to different intensities of exercise in sedentary young women. Obesit Med. 2016;1:15-20. [DOI:10.1016/j.obmed.2015.12.002]
- Moraes C, Leal VO, Marinho SM, et al. Resistance exercise training does not affect plasma irisin levels of hemodialysis patients. Horm Metab Res. 2013;45(12):900-4. [DOI:10.1055/s-0033-1354402]
- Tsuchiya Y, Ando D, Takamatsu K, Goto K. Resistance exercise induces a greater irisin response than endurance exercise. Metab Clin Experiment. 2015;64(9):1042-50. [DOI:10.1016/j.metabol.2015.05.010]
- 39. Kim H-j, So B, Choi M, Kang D, Song W. Resistance exercise training increases the expression of irisin concomitant with improvement of muscle function in aging mice and humans. Experiment Gerontol. 2015;70(1):11-7. [DOI:10.1016/j.exger.2015.07.006]
- Fox J, Rioux BV, Goulet EDB, et al. Effect of an acute exercise bout on immediate post-exercise irisin concentration in adults: A meta-analysis. Scandinavian J Med Sci Sports. 2018;28(1):16-28. [DOI:10.1111/sms.12904]

- Norheim F, Langleite TM, Hjorth M, et al. The effects of acute and chronic exercise on PGC-1α, irisin and browning of subcutaneous adipose tissue in humans. FEBS J. 2014;281(3):739-49. [DOI:10.1111/febs.12619]
- 42. Leung PS. The potential of irisin as a therapeutic for diabetes. Future Science; 2017. p. 18-26.
- 43. Piccirillo R. Exercise-induced myokines with therapeutic potential for muscle wasting. Front Physiol. 2019;10:287. [DOI:10.3389/fphys.2019.00287]
- Xie T, Leung PS. Roles of FGF21 and irisin in obesity-related diabetes and pancreatic diseases. J Pancreatol. 2020;3(1):29-34. [DOI:10.1097/JP9.000000000000039]
- Roca-Rivada A, Castelao C, Senin LL, et al. FNDC5/irisin is not only a myokine but also an adipokine. PloS one. 2013;8(4):56-63. [DOI:10.1371/journal.pone.0060563]
- 46. Zhou X, Xu M, Bryant JL, Ma J, Xu X. Exercise-induced myokine FNDC5/irisin functions in cardiovascular protection and intracerebral retrieval of synaptic plasticity. Cell Biosci. 2019;9(1):32. [DOI:10.1186/s13578-019-0294y]
- 47. Abdi A, Mehrabani J, Nordvall M, Wong A, Fallah A, Bagheri R. Effects of concurrent training on irisin and fibronectin type-III domain containing 5 (FNDC5) expression in visceral adipose tissue in type-2 diabetic rats. Arch Physiol Biochem. 2020;1(1):1-6. [DOI:10.1080/13813455.2020.1716018]
- Kaisanlahti A, Glumoff T. Browning of white fat: agents and implications for beige adipose tissue to type 2 diabetes. J Physiol Biochem. 2019;75(1):1-10. [DOI:10.1007/s13105-018-0658-5]
- 49. Fyfe JJ, Bishop DJ, Bartlett JD, et al. Enhanced skeletal muscle ribosome biogenesis, yet attenuated mTORC1 and ribosome biogenesisfollowing related signalling, short-term concurrent versus single-mode resistance Sci Reports. 2018;8(1):560. training. [DOI:10.1038/s41598-017-18887-6]
- Garneau L, Parsons SA, Smith SR, Mulvihill EE, Sparks LM, Aguer C. Plasma myokine concentrations after acute exercise in non-obese and obese sedentary women. Front Physiol. 2020;11(18). [DOI:10.3389/fphys.2020.00018]

How to Cite This Article:

Motahari Rad M, Bijeh N, Attarzadeh Hosseini R, Raouf Saeb A, The Impact of Different Modes of Exercise Training on Irisin: A Systematic Review and Meta-Analysis Research, J Adv Med Biomed Res. 2021; 29(134): 125-138

Download citation:

BibTeX | RIS | EndNote | Medlars | ProCite | Reference Manager | RefWorks

Send citation to: <u>Mendeley</u> 2 <u>Zotero</u> RefWorks <u>RefWorks</u>