

# Calcium phosphate bioceramics for improved angiogenesis

9

Farzad Kermani<sup>a</sup>, Saeid Kargozar<sup>b</sup>, Sergey V. Dorozhkin<sup>c</sup> and Sahar Mollazadeh<sup>a</sup>

<sup>a</sup>Department of Materials Engineering, Faculty of Engineering, Ferdowsi University of Mashhad (FUM), Mashhad, Iran, <sup>b</sup>Tissue Engineering Research Group (TERG), Department of Anatomy and Cell Biology, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran, <sup>c</sup>Kudrinskaja Square 1-155, Moscow, Russia

## 9.1 Introduction

During the last decades, there has been an excessive need for artificial bone tissue substitutes across the globe. The main reasons for such demand are associated with the increased aging population, obesity, cancers, accidents, and congenital genetic abnormalities. Accordingly, several attempts have been made to design and develop potent replacements capable of accelerating bone tissue healing. Previously, the specific and strict criteria have been defined by governmental agencies and regulatory bodies (e.g., Food and Drug Administration [FDA] and International Organization for Standardization [ISO], respectively) for any substance that should be implanted into the body. From a clinical point of view, the biocompatibility of materials is of utmost importance and defined through a series of well-established assays (e.g., cytotoxicity, hemocompatibility, etc.). However, biomaterials designed to repair and reconstruct damaged bones should meet other criteria, both structurally and functionally. In general, osteoconduction, osteoinduction, and osteointegration are stated as ideal biological properties of bone substitutes. Osteoconductive materials provide a proper substrate for bone growth on their surface. Osteoinductive materials can induce osteogenesis (new bone formation) through the recruitment of osteogenic cells to the injured region. Osteointegration means the ability of an exogenous material to incorporate within the bone and strongly attach to the surrounding bone [1].

Calcium phosphates (CaPs) represent versatile materials for biomedical applications, including bone tissue reconstruction. Hydroxyapatite (HAp) and beta-tricalcium phosphate ( $\beta$ -TCP) are two well-known members of the CaPs bioceramics family. It has been shown that CaPs and their constructs possess mechanical properties (e.g., compressive strengths) closely in the range of human bones. Still, the low strength, high brittleness, as well as limited bioactivities prohibit the extensive usage of CaPs bioceramics in many load-bearing applications [2,3]. Another disadvantage of CaPs is attributed to their poor degradation rate that inhibits bone ingrowth in implants and suppresses osteogenesis [3]. CaPs, predominantly in the form of ion-substituted

HAp, form the inorganic phase of bone tissues; therefore, they have been extensively investigated for bone repair and regeneration. Though, the solubility of CaPs is probably one of the most important characteristics that should be controlled as it may interface with the tissue healing process. Prior experiments have shown that CaPs can properly support the adhesion and proliferation of bone-forming cells (e.g., osteoblasts) *in vitro* and *in vivo* [4]. Thus, CaPs bioceramics (e.g., HAp) are generally recognized as osteoconductive biomaterials [5]. Although the osteoinductive properties of CaPs are attributed to their microstructural surface properties (grain size, microporosity, surface roughness, and specific surface area), this feature is not well-identified via *in vivo* animal studies [6,7]. In addition to improving osteogenesis, specific types of CaPs bioceramics showed the capability of inducing neovessel formation (angiogenesis). It has currently become a scientific fact that materials having proangiogenic activity can be more effective for accelerating wound healing as regards newly formed blood vessels are responsible for providing nutrients and oxygens to cells and removing waste metabolites from the injured site [8,9]. Respecting this critical issue, we aim to provide substantial information on the proangiogenic potential of CaPs bioceramics and its importance in developing of novel therapies for bone tissue engineering.

## 9.2 CaPs bioceramics: An overview

CaPs bioceramics have a long history in medicine with huge numbers of successfully performed clinical studies. Indeed, CaPs represent the main inorganic constituents of bone tissues in the body [10]. Hence, they are extensively applied for managing hard tissue defects. However, recent publications have declared specific types of CaPs could be used for treating soft tissue injuries [11]. These substances exhibit remarkable compatibility with the living systems (cells, tissues, and organs) and their by-products cause no adverse effects in the human body [10,12]. CaPs are generally bioresorbable compounds that can be replaced with natural tissue during the bone healing process. The dissolution rate of CaPs can greatly affect their biological functions (e.g., osteogenesis); therefore, a series of physico-chemical methods have been utilized to tune CaPs bioceramics properties. One of the most promising approaches is to dope metallic and nonmetallic elements to the CaPs network for generating potent substances in terms of osteogenesis and angiogenesis. For instance, lithium (Li), zinc (Zn), strontium (Sr), and magnesium (Mn) doped CaPs have exhibited improved osteogenesis while doping of copper (Cu) and cobalt (Co) to CaPs make them proangiogenic materials [13, 14]. It worth mentioning that some elements (e.g., boron (B) and silicon (Si)) can simultaneously improve osteogenic and angiogenic properties of CaPs [15]. Surface functionalization of CaPs is recognized as another approach applied for improving their biological activities. In this regard, surface properties of CaPs, including surface charge and roughness, can be optimized by physico-chemical modification methods. Experimental data have clarified that surface-modified CaPs can facilitate cellular interactions and improve cell adhesion, proliferation, and differentiation [16]. In addition to affecting biological properties, the defects could change the

physico-chemical properties of CaPs, including band-gap, surface charge, surface area, and ions release/uptake [17,18].

The inherent properties, including crystallographic and thermodynamic parameters, of commonly used CaPs in tissue engineering approaches are shown in Table 9.1. As it can be seen in Table 9.1, the ratio of Ca/P strongly influences the crystallographic parameters. Structural features and atomic distribution of CaPs were proven to be influential in determining their biological performance.

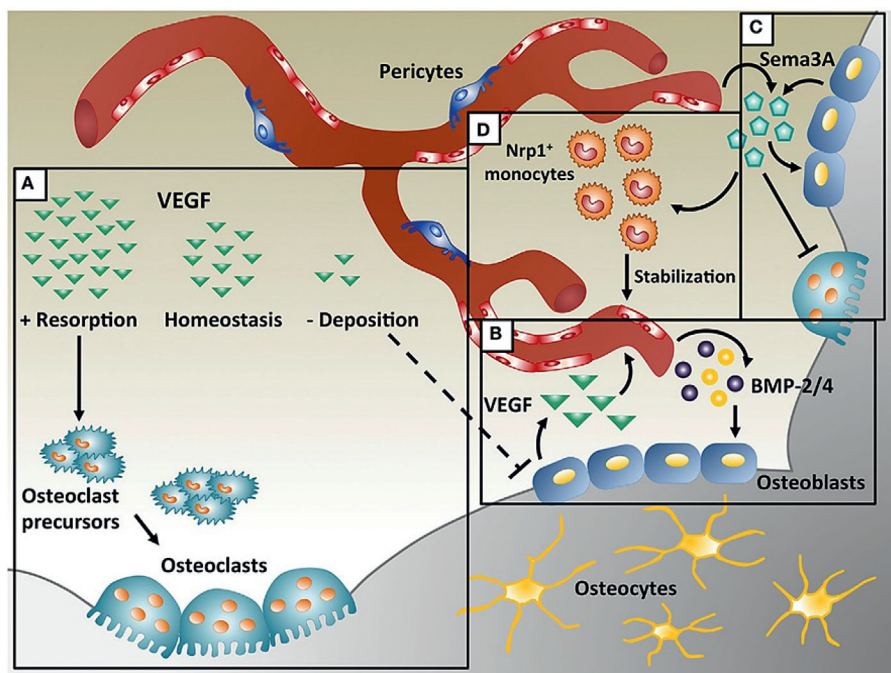
### 9.3 Clinical significance of angiogenesis in bone regeneration

In the body, blood vessels are responsible for providing oxygen and nutrients to cells and tissues as well as removing waste materials. The vasculature is obviously found in connective tissues like the bone [19]. The vasculature network of the bone is predominantly formed via angiogenesis. Developmental biology studies specify that the bone formation and vascularization processes concomitantly take place and can proceed through either endochondral ossification or intramembranous ossification. The vast majority of bones, including long bones (e.g., the femur), are generated by the endochondral ossification. On the other side, flat bones (e.g., the skull bones) are formed by using the intramembranous ossification in which mesenchymal cells are directly differentiated into osteoblasts. It should be mentioned that flat bones consist of a layer of compact bone interspersed with bone marrow [20]. In addition to skeletal development, the formation of new blood vessels is crucial during fracture healing [21]. Immediately after the damage, the volume of the vascular bed of bone tissue significantly increases due to the vasodilation process. This is followed by the formation of fracture hematoma, serving as a template for the development of the provisional, vascular bone callus [22]. The newly formed blood vessels may create other benefits for the bone healing process as to their angiocrine function (secreting paracrine signals) that is useful for coordinating of growth and differentiation of osteogenic progenitor cells [23].

Experimental data indicates that diverse molecular mediators (e.g., growth factors and cytokines) govern highly orchestrated interactions between different cells (macrophages, mesenchymal stem cells, etc.) to form new mineralized bone tissue [24,25]. Among different molecular mediators, vascular endothelial growth factor-A (VEGF-A) is known as a master regulator of bone development and regeneration (Fig. 9.1) [26]. In fact, these molecules can promote the migration and proliferation of endothelial cells (ECs) as well as stimulate the osteogenesis process through the regulation of osteogenic GFs [27,28]. During bone tissue repair, VEGF is generally produced by osteoblasts and enhances the ECs' migration and proliferation [29]. It should be mentioned that the physiological level of VEGF is strictly regulated as its low values may interrupt osteoblast differentiation, and while its high values increase osteoclast recruitment, leading to bone resorption [22].

**Table 9.1** A summary of crystallographic and thermodynamic parameters of calcium phosphates (CaPs) [12].

Compound	Formula	Ca/P ratio	Space group	Unit cell parameters (Å and °)	Z	Density (g.cm <sup>-3</sup> )	pH stability in aqueous solution (298.15 K)	Thermodynamic parameters (298.15 K)		
								H° kJ/mol	S° J/(mol·K)	C <sub>p</sub>
Monocalcium phosphate monohydrate (MCPM)	Ca(H <sub>2</sub> PO <sub>4</sub> ) <sub>2</sub> ·H <sub>2</sub> O	0.5	Triclinic Pī	a = 5.6261(5), b = 11.889(2), c = 6.4731(8) α = 98.633(6) β = 118.262(6) γ = 83.344(6)	2	2.23	0.0 ± 2.0	-3410.0	259.8	165.0 + 318T
Monocalcium phosphate anhydrate (MCPA)	Ca(H <sub>2</sub> PO <sub>4</sub> ) <sub>2</sub>	0.5	Triclinic Pī	a = 7.5577(5), b = 8.2531(6), c = 5.5504(3) α = 109.87(1), β = 93.68(1), γ = 109.15(1)	2	2.58	–	-3115.0	189.5	259.8
Dicalcium phosphate dihydrate (DCPD, “brushite”)	CaHPO <sub>4</sub> ·2H <sub>2</sub> O	1	Monoclinic Ia	a = 5.812(2), b = 15.180(3), c = 6.239(2) β = 116.42(3)	4	2.32	2.0 ± 6.0	-2421.2	189.5	225.9 + 55T
Dicalcium phosphate anhydrate (DCPA, “monetite”)	CaHPO <sub>4</sub>	1	Triclinic Pī	a = 6.910(1), b = 6.627(2), c = 6.998(2) α = 96.34(2), β = 103.82(2), γ = 88.33(2)	4	2.89	–	-1814.4	111.4	138.4 + 55T
Octacalcium phosphate (OCP)	Ca <sub>8</sub> (HPO <sub>4</sub> ) <sub>2</sub> (PO <sub>4</sub> ) <sub>4</sub> ·5H <sub>2</sub> O	1.33	Triclinic Pī	a = 19.692(4), b = 9.523(2), c = 6.835(2) α = 90.15(2), β = 92.54(2), γ = 108.65(1)	1	2.61	5.5–7.0	-13375.2	878.22	883.1
α-Tricalcium phosphate (α-TCP)	α-Ca <sub>3</sub> (PO <sub>4</sub> ) <sub>2</sub>	1.5	Monoclinic P2 <sub>1</sub> /a	a = 12.887(2), b = 27.280(4), c = 15.219(2) β = 126.20(1)	24	2.86	–	-4109.9	240.9	231.6
β-Tricalcium phosphate (β-TCP)	β-Ca <sub>3</sub> (PO <sub>4</sub> ) <sub>2</sub>	1.5	Rhombohedral R3Ch	a = b = 10.439(1), c = 37.375(6) γ = 120	21	3.07	–	-4120.8	236.0	227.8
Amorphous calcium phosphate (ACP)	Ca <sub>x</sub> (PO <sub>4</sub> ) <sub>y</sub> ·nH <sub>2</sub> O	1.2–2.2	–	–	–	–	–	–	–	–
Calcium-deficient hydroxyapatite (CDHA)	Ca <sub>10-x</sub> (HPO <sub>4</sub> ) <sub>x</sub> (PO <sub>4</sub> ) <sub>6-x</sub> ·2-x (0 < x < 1)	1.5–1.67	–	–	–	–	6.5–9.5	–	–	–
Hydroxyapatite (HA)	Ca <sub>10</sub> (PO <sub>4</sub> ) <sub>6</sub> O <sub>2</sub>	1.67	Monoclinic P2 <sub>1</sub> /b	a = 9.84214(8), b = 2a, c = 6.8814(7) γ = 120	4	3.16	9.5–12	–	–	–
			Hexagonal P6 <sub>3</sub> /m	a = b = 9.4302(5), c = 6.8911(2) γ = 120	2			-13477.0	780.7	963.5 + 108.2T
Tetracalcium phosphate (TTCP)	Ca <sub>4</sub> (PO <sub>4</sub> ) <sub>2</sub> O	2	Monoclinic P2 <sub>1</sub>	a = 7.023(1), b = 11.986(4), c = 9.473(2) β = 90.90(1)	4	3.05	–	-4940.6	275.5	271.5



**Figure 9.1** Schematic representation displaying the coupling of angiogenesis and osteogenesis during intramembranous ossification. (A) Vascular endothelial growth factor (VEGF) at physiological levels guarantees bone homeostasis. (B) During bone repair, VEGF secreted by osteoblasts promotes the migration and proliferation of endothelial cells (ECs). In turn, ECs secrete osteogenic factors (e.g., bone morphogenetic protein (BMP)-2 and -4) and thereby support osteoblast differentiation. (C) VEGF can regulate the expression of semaphorin 3A (Sema3A) in ECs in a dose-dependent manner; Sema3A suppresses osteoclast differentiation and induces bone deposition. (D) Sema3A is also identified as the main mediator for the recruitment of neuropilin 1-expressing (Nrp1 +) monocytes, which improve vessel stabilization. Reproduced from [22].

## 9.4 CaPs bioceramics for improved angiogenesis

Tissue healing is a complicated process in which well-orchestrated biological events happen for obtaining a timely repair. In the case of bone tissue, osteoconductivity, osteoinductivity and osseointegration are regarded as three main features defined for any ideal substitute to achieve a perfect healing process [30]. Previously reported research indicates that CaPs (HAp, TCP, biphasic CaPs (BCP = HAp + TCP), etc.) are among the osteoconductive biomaterials [4]. In addition, CaPs are known suitable substances for bone tissue engineering due to their capability of enhancing osteogenesis. It is well-recognized that failing to provide sufficient nutrients and oxygen to cells at damaged sites may result in failure in tissue transplantation. Therefore, numerous attempts have been made to produce CaPs with higher angiogenic capacity in the last decade.

Currently, the proangiogenic capability of CaPs-based tissue substitutes takes into account the meantime of designing and preparing the final products. Different types and forms of CaPs have shown the ability to improve neovascularization [31]. Several parameters can determine the proangiogenic capacity of CaPs, including their chemical formulation and physical properties. In this regard, the role of physical characteristics was stated as a key determinant in the angiogenic potential of CaPs ceramics. For illustration, TCP with submicron surface topography could more effectively stimulate M1 to M2 macrophage polarization and thereby enhance the secretion of proangiogenic growth factor and chemokine TGF- $\beta$  and CCL18 as compared with the samples having micron-scale topography [32]. The in vivo results confirmed that the intramuscular implantation of TCPs with submicron surface topography into adult male beagles could increase typical M2 macrophage markers (e.g., IL-10), leading to enhance blood vessel formation and improved bone formation. Prior studies have clarified that positive effects of CaPs on new blood vessel formation can even be observed after making composite constructs with them. In this sense, octacalcium phosphate (OCP,  $\text{Ca}_8\text{H}_2(\text{PO}_4)_6 \cdot 5\text{H}_2\text{O}$ ) included in gelatin could trigger the capillary-like tube formation in human umbilical vein endothelial cells (HUVECs) in a dose-dependent manner [33]. Some innovative approaches were developed over the last decades to improve the proangiogenic potential of CaPs ceramic, including doping of proangiogenic elements in their chemical composition as well as surface modification approaches. These strategies are introduced and discussed in the following sections based on the recent publications.

#### **9.4.1 Doped CaPs bioceramics for promoted angiogenesis**

There are several trace elements (e.g., copper) in the human body that are needed for mediating a wide range of biochemical reactions. Previously, biological impacts of these metallic elements have been entirely studied through in vitro or in vivo assays [34,35]. Improving cell proliferation, providing antibacterial and anti-inflammatory properties, as well as promoting angiogenesis are some of the most well-known biological features rendered by administering trace elements [36]. Therefore, adding trace elements to the structure of CaPs ceramics has become a routine approach in the biomedical setting to positively affect the tissue repair and regeneration processes [37]. In fact, doped-CaPs can be served as drug delivery vehicles for providing a sustained release of therapeutic ions at damaged sites. Concerning neovascularization, specific kinds of trace metallic elements (e.g., copper and cobalt) may be incorporated into CaPs structure to promote angiogenesis either in vitro or in vivo (Table 9.2).

The introduction of proangiogenic metallic dopants to CaPs ceramics was confirmed as a safe and efficient strategy in favor of tissue engineering. It should be emphasized that this process may alter the physico-chemical, mechanical and biological properties of CaPs [38]. Silicon (Si), copper (Cu), cobalt (Co), and boron (B) are among the most well-studied elements for inducing angiogenesis [39]; they could be easily incorporated into CaPs structure and make a proangiogenic substitute for accelerating wound healing. Silicon is generally recognized as a semi-metal material (metalloid) with some critical functions in the human body. Silicate ions ( $\text{SiO}_4^{4-}$ ) can play a direct

**Table 9.2** A summary of calcium phosphates (CaPs) ceramics doped with proangiogenic elements for potential use in bone tissue engineering.

CaPs ceramics	Dopants	Remarks	Ref
60 wt% TCP, 25 wt% dicalcium phosphate anhydrous (DCPA), 10 wt% HAp nanoparticles, and 5 wt% calcium carbonate (CC)	Copper at dosages of 0.01, 0.05, 0.1 and 0.5 wt%	The 0.01 wt% Cu-doped enhanced: - The proliferation of bone marrow mouse bone marrow stromal cells (mBMSCs) - The expression of osteogenic differentiation-related genes (collagen I and osteocalcin) and proteins (ALP and collagen I). - The proliferation of HUVECs - The expression of angiogenesis-related genes (eNOs, VEGF and bFGF), proteins (VEGF and NO) - <i>in vitro</i> tube formation of HUVECs	[36]
HAp/ $\beta$ -TCP with a ratio of 1.67	Silicon at dosages of 0.1, 0.4 mol %	The 0.1 mol% Si <sup>4+</sup> doped samples could: - Increase <i>in vitro</i> bone nodule formation - improve cell migration rate	[45]
Brushite cement (Ca/P ratio of ~ 1.5) composited with silk	Silicon and zinc at dosages of 0.5 and 0.25 wt %, respectively	The Si/Zn doped samples could: - Increase the nitric oxide (NO) secretion by endothelial cells - Overexpress hypoxia-inducible factor 1 alpha (HIF-1 $\alpha$ ) - Enhance tube formation	[48]
CaP-coated poly (D,L-lactic acid) (PLA)	Cobalt at dosages of 0.1 and 20 $\mu$ M	– 0.1 $\mu$ M Co <sup>2+</sup> doped CaPs could: - Improve the upregulation of HIF-1 $\alpha$ <i>in vitro</i> - Generate greater blood vessel area and higher numbers of large blood vessels <i>in vivo</i>	[58]
$\beta$ -TCP/HAp scaffolds containing growth differentiation factor-5 (GDF-5)-loaded poly(lactide-co-glycolide) (PLGA) microspheres	Copper and zinc at dosages of 0.005, 0.02, and 0.05 M	The Cu/Zn-doped composites could: - Improve the proliferation of BMSCs and vascular endothelial cells - Enhance the VEGF secretion	[76]

role in the mineralization of bones during the later stages of calcification [40]. The proangiogenic activity of silicate ions has been previously demonstrated; they were extensively applied in tissue engineering strategies for obtaining accelerated wound healing. The presence of silicates can induce neovessel formation by upregulating endothelial nitric oxide synthase (eNOS), leading to increased production of a series of proangiogenic growth factors (VEGF, bFGF, and TGF- $\beta$ ) [41]. In addition, these ions may increase the  $\text{Ca}^{2+}$  levels in cells and thereby affect the proliferation and motility of vascular ECs in favor of sprout outgrowth of new blood vessels [42–44]. Accordingly, Si-doped CaPs were developed, and reported data indicated superior biological properties of these materials for tissue repair and regeneration applications as compared to their un-doped counterparts [45]. As previously stated by Prof. Bohner, apart from passive mechanisms (change in the grain size and protein conformation at the material surface) that describe biological effects of Si substitution in materials, an active mechanism, i.e., Si release, is responsible for positive changes in cellular behaviors [46,47]. The proangiogenic activities of Si-doped CaPs were even observed after adding to a polymeric substrate both in vitro and in vivo, holding promises in bone tissue engineering [48].

Copper (Cu) is known as an essential cofactor in humans and shows desirable therapeutic properties for managing different types of disorders and diseases. On this matter, the antibacterial activity of Cu was recently reviewed, and its usefulness in orthopedic applications has been highlighted [49]. On the other side, Cu was approved for its biological effectiveness in activating several growth factors, cytokines, and molecules involved in advancing the angiogenesis process, including VEGF, FGF-1 and -2, fibronectin, angiogenin [50].  $\text{Cu}^+/\text{Cu}^{2+}$  ions were demonstrated to be effective in the initiation, maturation, and regulation of blood vessel formation (ECM remodeling). Cu may promote angiogenesis via two specific signaling pathways of (I) the hypoxia-inducible HIF-1 and (II) the mitogen-activated protein kinase (MAPK) pathways [51]. Cu-containing CaPs bioceramics (e.g., Cu-doped HAp and  $\beta$ -TCP) have been successfully synthesized and demonstrated for their ability to promoting neovessel formation [35,52,53]. For instance, an improved angiogenesis was observed after the incubation of HUVECs with 0.01 wt% and 0.05 wt% Cu-doped calcium phosphate cement [36]. Additionally, the fabrication of 3D scaffolds from Cu-doped ceramics has also been evaluated for potential use in bone tissue engineering strategies [54,55]. As cutting-edge research, designing and developing of 3D printed scaffolds made of Cu-doped CaPs may open new horizons in the orthopedic setting with huge opportunities.

Cobalt is another proangiogenic trace element in the human body, which is normally found at the center of vitamin B12 and some other co-enzymes (named cobalamins). Although cobalt is generally recognized as a toxic element, it can be used for medical purposes at very low concentrations without any adverse effects. Divalent cobalt ions ( $\text{Co}^{2+}$ ) could stimulate neovascularization through specific signaling pathways; for example, they mimic hypoxia conditions via the stabilization of hypoxia-inducible factor-1 alpha (HIF-1 $\alpha$ ), which lead to the upregulation of proangiogenic factors VEGF and bFGF [56,57]. Adding cobalt into different types of CaPs bioceramics was previously investigated, and reported data have been quite promising in terms of improved angiogenesis capacity [58]. As an illustration,  $\beta$ -TCP powders doped



with different concentrations of cobalt (0, 2, 5 mol%) showed the ability to increase VEGF expression in human bone marrow mesenchymal stem cells (HBMSCs) as well as the capacity of the tube-like structure formation in HUVECs. The findings clarified that the substitution of cobalt with calcium in the  $\beta$ -TCP structure might also lead to suppressing the phase transition from  $\beta$ - to  $\alpha$ -TCP [59]. Cobalt-substituted HAp powders are another successful example of proangiogenic CaPs ceramics; 0.33% (w/w)  $\text{Co}^{2+}$  doping to HAp network resulted in stabilizing HIF-1 $\alpha$  and consequently enhanced production of VEGF in human osteoblast-like cells (MG-63) [60]. There are several studies indicating the effectiveness of three-dimensional (3D) constructs made of  $\text{Co}^{2+}$ -substituted CaPs ceramics in bone tissue engineering [61]. Still, there is a scientific gap in designing and developing 3D printed constructs of cobalt-substituted CaPs bioceramics for potential use in hard tissue engineering. Still, the main concern on using  $\text{Co}^{2+}$ -doped CaPs bioceramics is associated with the risk of toxic effects of cobalt for living systems. Accordingly, cautions should be taken in the case of in vivo implantation of cobalt-containing CaPs. In addition, the sustained release of  $\text{Co}^{2+}$  ions from CaPs-based scaffolds should be carefully considered as a critical factor in determining biological outcomes.

Boron (B) is a nonmetal micronutrient in the human body that plays varying vitally important roles in a wide range of biological functions (e.g., bone metabolism). The level of this trace element in human bone tissue is about 0.90 ppm and is regarded as a major factor in preventing skeletal loss and protecting the organism from the development of osteoporosis [62,63]. Considering its biological properties, boric acid ( $\text{H}_3\text{BO}_3$ ) at low concentrations can activate the MAPK signaling pathway, leading to improved cell growth and proliferation [64]. Furthermore,  $\text{H}_3\text{BO}_3$  can stimulate the translation of mRNAs encoding proangiogenic proteins, including VEGF and transforming growth factor  $\beta$  (TGF- $\beta$ ) [65]. Therefore, this trace element has been considered in proangiogenic strategies for years; B-containing biomaterials were developed as new types of tissue substitutes capable of accelerating wound healing [66,67]. The substitution of borate groups into CaPs ceramics may happen on  $\text{PO}_4$  and OH sites, predominately the first [68]. B-doped biphasic HAp/ $\beta$ -TCP samples were previously synthesized and characterized for potential use in bone tissue engineering [69]. Moreover, the in vitro osteogenic capacity of composite scaffolds based on B-doped HAp and chitosan was previously reported [70]. However, there is a noticeable shortcoming in experimental studies on utilizing and investigating B-substituted CaPs bioceramics for improved angiogenesis. It is worth noting that the incorporation of nanoplateforms of B (e.g., nanotubes) into CaPs bioceramics was also evaluated, and reported data has been promising for stimulated new bone formation [71]. Still, the proangiogenic activity of such compositions has not well-studied either by in vitro or in vivo assays.

In addition to the elements mentioned above, a couple of chemicals with proangiogenic capacity were also substituted into the CaPs network for promoting their inherent biological properties in terms of neovascularization [72,73]. Zinc (Zn) is among the most promising elements exhibiting proangiogenic activities through upregulation of relevant growth factors like VEGF and FGF [74,75]. This element, either alone or in combination with other elements, can be easily incorporated into CaPs structure and applied for tissue engineering applications [34]. It should be stated that the proangiogenic

potential of most of these elements in CaPs is dose-dependent. All in all, although the primary results of CaPs bioceramics substituted with proangiogenic dopants have been quite promising in terms of tissue engineering applications, some critical points remain to be solved. For example, achieving a sustained release of dopants during a long period seems necessary with regard to the tissue repair timeline. This will be even more complicated if doped CaPs apply for managing injuries and damages related to other tissues (e.g., dental). The second limitation may correlate to the necessary amounts of each proangiogenic dopant in CaPs structure. High concentrations of some trace elements could cause cellular and molecular damages and thereby hinder the healing process. For example, although cobalt may promote new blood vessel formation, cautions should be taken into accounts as it is a toxic element at specific dosages for the living system.

#### **9.4.2 Functionalized CaPs for induced angiogenesis**

Experimental studies have demonstrated that surface properties, including chemical composition and surface structure, could play an important role in the biological properties of biomaterials [77]. In addition, the surface can be considered an appropriate place to graft bioactive molecules and improve their biological performance, such as angiogenesis capability [78]. Up to now, several surface modification techniques were well developed and applied to generate more suitable substitutes for tissue regenerations applications, including grafting, coating, and so on [79]. Physical adsorption of a proangiogenic GF (i.e., VEGF) to CaPs bone cement Bio cement D was previously reported effective for activating angiogenesis by sustained delivery of VEGF [80]. The importance of small molecules and oligonucleotides is clearly understood in inducing or suppressing biological reactions in mammalian cells [81]. There are some interesting reports in the literature in which CaPs bioceramics have been armored with these therapeutic molecules and applied for obtaining accelerated tissue healing. For instance, the 3R02 bivalent DNA aptamer was immobilized onto HAp particles by using 3-aminopropyltriethoxysilane (APTES), and obtained in vivo data indicated that the particles are suitable candidates for enhancing angiogenesis and osteogenesis as regards their ability to capture higher VEGF protein and enhance the growth of HUVECs compared to pristine counterparts [82]. It should be highlighted that the use of linkers (e.g., peptide linkers) maybe as a beneficial for the grafting and immobilizing of different molecules onto CaPs surface at the same time in defined ratios [83]. Prior experiments have well-proved that specific sorts of plant extracts can activate molecular signaling pathways involved in angiogenesis in a dose-dependent manner. This issue can be acknowledged as a unique opportunity for designing novel proangiogenic CaPs in a green route.

### **9.5 CaPs bioceramics for delivery of proangiogenic bioactive molecules**

CaPs bioceramic are generally identified as suitable vehicles for loading and delivery of a wide range of chemicals and bioactive molecules (proteins, nucleic acids, and so

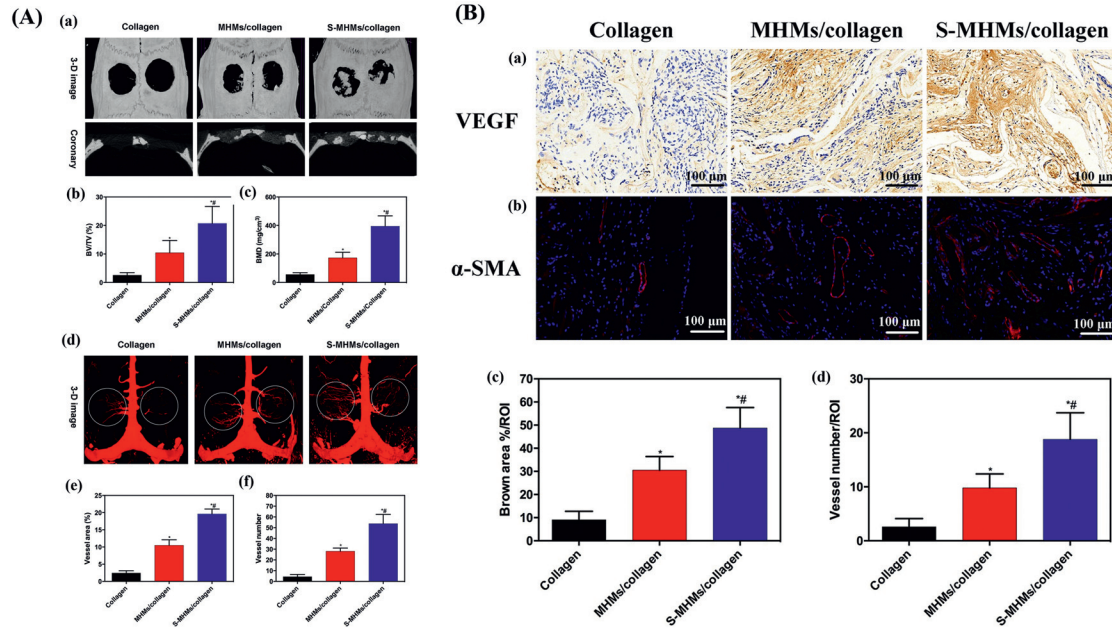
on) [84]. In fact, CaPs nanoparticles can provide an increase in the local drug levels in the desired areas and decrease the risks of systemic toxicity [85]. Additionally, achieving a prolonged drug release is feasible by CaPs ceramics, ranging from insoluble, hydrophobic or hydrophilic drugs. This feasibly is also practicable in the case of CaPs-based composites [86]. The possibility to make stimuli-responsive delivery systems by CaPs particles is considered outstanding merit for their applications in biomedicine. For example, thermo- and pH-sensitive CaPs ceramics were previously developed and successfully applied for biomedical approaches [87,88].

Focusing on the angiogenesis issue, CaPs ceramics in different shapes and forms have been investigated for loading and delivery of proangiogenic cargoes [89]. For instance, mesoporous HAp microspheres were successfully utilized as a sustained release system for delivery of simvastatin to improve osteogenesis and angiogenesis and consequently accelerated bone regeneration (Fig. 9.2) [90]. Furthermore, researchers were succeeded in developing injectable CaPs foams based on TCP with the capability of loading and delivery of Pitavastatin as an osteogenic and angiogenic agent [91]. This system could serve as a delivery platform for the controlled release of Pitavastatin and enhanced *in vitro* mineralization and vascularization in a dose-dependent manner. It is worth mentioning that *in vivo* degradation of CaPs ceramics can be regarded as a beneficial parameter in providing a sustained release profile of proangiogenic molecules. On this matter, TGF- $\beta$ 1 adsorbed to  $\beta$ -TCP was previously reported to have the ability to induce neovascularization and enhance bone formation [92].

It should be mentioned that the pore size and porosity of CaPs-based scaffolds were previously introduced as important determinants affecting bone formation and angiogenesis [93]. Scaffold porosity was reported as a more dominant player in promoting bone formation and neovascularization in comparison with the pore size. Though, the release rate and release fraction of osteogenic and proangiogenic growth factors were mostly controlled by the scaffold's pore size. In addition to conventionally fabricated constructs, 3D printed CaPs bioceramic implants are also utilized for loading and delivery of proangiogenic growth factors (e.g., VEGF) with promising reported data on their effectiveness in promoted neovascularization and wound healing [94,95]. Still, the current knowledge on this issue should be extended by performing more experimental studies, especially those are associated with *in vivo* studies.

## 9.6 Summary and future perspectives

Calcium phosphates (CaPs) are among the most promising materials for managing broad ranges of skeleton systems. Importantly, the main inorganic phase of native bone is made of calcium and phosphates elements, predominantly in the form of ion-substituted Ca-deficient hydroxyapatite crystals. That is why CaPs bioceramics exhibit inherent properties in favor of bone tissue regeneration, including excellent biocompatibility, bioactivity and osteoconductivity. In recent decades, understanding the real capacity of CaPs ceramics in stimulating angiogenesis, that is, new blood vessel formation has been one of the hottest topics in materials science and bioengineering fields. It is well-known that the degree of bone repair directly depends on



**Figure 9.2** (A) Three dimensional (3-D) and coronary views of reconstructed calvaria bone treated by collagen, mesoporous HAp microspheres (MHMs)/collagen, simvastatin-loaded MHMs (a); bone mineral density (BMD), and bone volume (BV)/total volume (TV) in the defect sites (b and c); newly formed blood vessels provided by 3-D reconstruction images; quantitative analysis of the new blood vessel area and number (e and f). (B) Immunohistochemistry evaluation of angiogenic markers in each group, including VEGF and  $\alpha$ -SMA. \*Comparison between collagen and other groups, #comparison between the MHMs/collagen and S-MHMs/collagen groups,  $p < .05$ ). Reproduced with permission from [90].

the number of new blood vessels in the damaged region. In fact, blood vessels are responsible for providing sufficient levels of oxygen and nutrients to cells and tissues and remove their waste metabolites from the affected site. Accordingly, the design and preparation of proangiogenic CaPs can be useful for obtaining accelerated bone tissue healing. Scientific evidence indicates both physical characteristics and the chemical composition may affect the angiogenic potential of CaPs. On this matter, surface topography is recognized as a primary parameter in the proangiogenic activity of CaPs bioceramics. Understanding the ion release mechanism from CaPs structure led to the emergence of the term “drug delivery systems” for these materials. Currently, a series of proangiogenic elements, mostly metallic ions such as  $\text{Cu}^{2+}$  and  $\text{Co}^{2+}$ , doped to the CaPs network for making substances with more angiogenic capacity. In addition, CaPs provide an outstanding opportunity for loading and delivery of proangiogenic drugs and chemicals into injured regions for boosting bone tissue healing. In this regard, plant-derived components and phytochemicals with proangiogenic activity (e.g., curcumin) may be suitable candidates for developing angiogenesis-inducing CaPs bioceramics. The fabrication of 3D printed CaPs scaffolds with the ability to stimulate angiogenesis is regarded a step-forward for bone tissue engineering applications. Still, more research, especially clinical trials, is required to perfectly design and conduct for revealing all the pros and cons of angiogenesis-inducing CaPs in managing bone tissue lesions.

## References

- [1] Böhner M. Calcium orthophosphates in medicine: from ceramics to calcium phosphate cements. *Injury* 2000;31:D37–47.
- [2] Zhang JT, Tancret F, Bouler JM. Fabrication and mechanical properties of calcium phosphate cements (CPC) for bone substitution. *Mater Sci Eng C* 2011;31(4):740–7.
- [3] Wu Q, Xu S, Wang X, Jia B, Han Y, Zhuang Y, et al. Complementary and synergistic effects on osteogenic and angiogenic properties of copper-incorporated silicocarnotite bioceramic: in vitro and in vivo studies. *Biomaterials* 2021;268:120553.
- [4] Bouler JM, Pilet P, Gauthier O, Verron E. Biphasic calcium phosphate ceramics for bone reconstruction: a review of biological response. *Acta Biomater* 2017;53:1–12.
- [5] LeGeros RZ. Properties of osteoconductive biomaterials: calcium phosphates. *Clin Orthop Relat Res* 2002;395:81–98.
- [6] Habibovic P, Yuan H, van der Valk CM, Meijer G, van Blitterswijk CA, Groot K. 3D microenvironment as essential element for osteoinduction by biomaterials. *Biomaterials* 2005;26(17):3565–75.
- [7] Yuan H, Fernandes H, Habibovic P, Boer J, Barradas Ana MC, de Ruiter Ad, et al. Osteoinductive ceramics as a synthetic alternative to autologous bone grafting. *Proc Natl Acad Sci* 2010;107(31):13614.
- [8] Kargozar S, Baino F, Hamzehlou S, Hill RG, Mozafari M. Bioactive glasses: sprouting angiogenesis in tissue engineering. *Trends Biotechnol* 2018;36(4):430–44.
- [9] Kargozar S, Francesco B, Hamzehlou S, Hamblin MR, Mozafari M. Nanotechnology for angiogenesis: opportunities and challenges. *Chem Soc Rev* 2020;49(14):5008–57.
- [10] Dorozhkin SV, Epple M. Biological and medical significance of calcium phosphates. *Angewandte Chemie Int Ed* 2002;41(17):3130–46.

- [11] Kargoazar S, Rajendra KS, Hae-Won K, Baimo F. “Hard” ceramics for “Soft” tissue engineering: paradox or opportunity? *Acta Biomater* 2020;115:1–28.
- [12] Habraken W, Habibovic P, Epple M, Bohner M. Calcium phosphates in biomedical applications: materials for the future? *Mater Today* 2016;19(2):69–87.
- [13] Šupová M. Substituted hydroxyapatites for biomedical applications: a review. *Ceram Int* 2015;41(8):9203–31.
- [14] Yu W, Sun T, Ding Z, Qi C, Zhao H, Chen F, et al. Copper-doped mesoporous hydroxyapatite microspheres synthesized by a microwave-hydrothermal method using creatine phosphate as an organic phosphorus source: application in drug delivery and enhanced bone regeneration. *J Mater Chem B* 2017;5(5):1039–52.
- [15] Kermani F, Mollazadeh S, Kargoazar S, Vahdati Khakhia J. Improved Osteogenesis and angiogenesis of theranostic ions doped calcium phosphates (CaPs) by a simple surface treatment process: a state-of-the-art study. *Mater Sci Eng C* 2021;124:112082.
- [16] Salamanca E, Pan Y-H, Tsai AI, Lin P-Y, Lin C-K, Huang H-M, et al. Enhancement of osteoblastic-like cell activity by glow discharge plasma surface modified hydroxyapatite/ $\beta$ -tricalcium phosphate bone substitute. *Materials*, 2017;10(12):1347.
- [17] Bystrov VS, Avakyan LA, Paramonova EV, Coutinho J. Sub-band gap absorption mechanisms involving oxygen vacancies in hydroxyapatite. *J Phys Chem C* 2019;123(8):4856–4865.
- [18] Peroos S, Du Z, de Leeuw NH. A computer modelling study of the uptake, structure and distribution of carbonate defects in hydroxy-apatite. *Biomaterials* 2006;27(9):2150–61.
- [19] Filipowska J, Tomaszewski KA, Niedźwiedzki L, Walocha JA, Niedźwiedzki T. The role of vasculature in bone development, regeneration and proper systemic functioning. *Angiogenesis* 2017;20(3):291–302.
- [20] Watson EC, Adams RH. Biology of bone: the vasculature of the skeletal system. *Cold Spring Harbor Persp Med* 2018;8(7):a031559.
- [21] Portal Núñez, S, Lozano D, and Esbrit P. Role of angiogenesis on bone formation. 2012.
- [22] Grosso A, Burger MG, Lunger A, Schaefer DJ, Banfi A, Di Maggio N. It takes two to tango: coupling of angiogenesis and osteogenesis for bone regeneration. *Front Bioeng Biotechnol* 2017;5:68.
- [23] Sivan U, De Angelis J, Kusumbe AP. Role of angiocrine signals in bone development, homeostasis and disease. *Open Biol* 2019;9(10):190144.
- [24] Pajarinen J, Lin T, Gibon E, Kohno Y, Maruyama M, Nathan K, et al. Mesenchymal stem cell-macrophage crosstalk and bone healing. *Biomaterials* 2019;196:80–9.
- [25] Leucht P, Lee S, Yim N. Wnt signaling and bone regeneration: can’t have one without the other. *Biomaterials* 2019;196:46–50.
- [26] Hu K, Olsen BR. The roles of vascular endothelial growth factor in bone repair and regeneration. *Bone* 2016;91:30–8.
- [27] Schipani E, Maes C, Carmeliet G, Semenza G. Regulation of osteogenesis-angiogenesis coupling by HIFs and VEGF. *J Bone Minere Res* 2009;24(8):1347–53.
- [28] Street J, Bao M, Guzman L, Bunting S, Peale FV Jr, Steinmetz H, et al. Vascular endothelial growth factor stimulates bone repair by promoting angiogenesis and bone turnover. *Proc Natl Acad Sci* 2002;99(15):9656–61.
- [29] Hu K, Olsen BR. Osteoblast-derived VEGF regulates osteoblast differentiation and bone formation during bone repair. *J Clin Invest* 2016;126(2):509–26.
- [30] Albrektsson T, Johansson C. Osteoinduction, osteoconduction and osseointegration. *Eur Spine J* 2001;10(2):S96–S101.
- [31] Huang G-J, Yu H-P, Wang X-L, Ning B-B, Gao J, Shi Y-Q, et al. Highly porous and elastic aerogel based on ultralong hydroxyapatite nanowires for high-

- performance bone regeneration and neovascularization. *J Mater Chem B* 2021;9(5): 1277–1287.
- [32] Duan R, Zhang Y, Dijk LV, Barbieri D, Jeroen van den Beucken JJJP, Yuan H, et al. Coupling between macrophage phenotype, angiogenesis and bone formation by calcium phosphates. *Mater Sci Eng C* 2021;122:111948.
- [33] Kurobane T, Shiwaku Y, Anada T, Hamai R, Tsuchiya K, Baba K, et al. Angiogenesis involvement by octacalcium phosphate-gelatin composite-driven bone regeneration in rat calvaria critical-sized defect. *Acta Biomater* 2019;88:514–26.
- [34] Fielding G, Bose S. SiO<sub>2</sub> and ZnO dopants in three-dimensionally printed tricalcium phosphate bone tissue engineering scaffolds enhance osteogenesis and angiogenesis in vivo. *Acta Biomater* 2013;9(11):9137–48.
- [35] Zhang J, Wu H, He F, Wu T, Zhou L, Ye J. Concentration-dependent osteogenic and angiogenic biological performances of calcium phosphate cement modified with copper ions. *Mater Sci Eng C* 2019;99:1199–212.
- [36] Lin Z, Cao Y, Zou J, Gao Y, Zhu F, Zheng X, et al. Improved osteogenesis and angiogenesis of a novel copper ions doped calcium phosphate cement. *Mater Sci Eng C* 2020;114:111032.
- [37] Bose S, Fielding G, Tarafder S, Bandyopadhyay A. Understanding of dopant-induced osteogenesis and angiogenesis in calcium phosphate ceramics. *Trends Biotechnol* 2013;31(10):594–605.
- [38] Bazin T, Magnaudeix A, Mayet R, Carles P, Julien I, Demourgues A, et al. Sintering and biocompatibility of copper-doped hydroxyapatite bioceramics. *Ceram Int* 2021;47(10):13644–54.
- [39] Kargozar S, Baino F, Hamzehlou S, Hill RG, Mozafari M. Bioactive glasses entering the mainstream. *Drug Discov Today* 2018;23(10):1700–4.
- [40] Pietak AM, Reid JW, Stott MJ, Sayer M. Silicon substitution in the calcium phosphate bioceramics. *Biomaterials* 2007;28(28):4023–32.
- [41] Li H, Chang J. Bioactive silicate materials stimulate angiogenesis in fibroblast and endothelial cell co-culture system through paracrine effect. *Acta Biomater* 2013;6(9):6981–91.
- [42] Li H, Xue K, Kong N, Liu K, Chang J. Silicate bioceramics enhanced vascularization and osteogenesis through stimulating interactions between endothelia cells and bone marrow stromal cells. *Biomaterials* 2014;35(12):3803–18.
- [43] Shi M, Zhou Y, Shao J, Chen Z, Song B, Chang J, et al. Stimulation of osteogenesis and angiogenesis of hBMSCs by delivering Si ions and functional drug from mesoporous silica nanospheres. *Acta Biomater* 2015;21:178–89.
- [44] Saghiri MA, Asatourian A, Orangi J, Sorenson CM, Sheibani N. Functional role of inorganic trace elements in angiogenesis—Part II: Cr, Si, Zn, Cu, and S. *Crit Rev Oncol/Hematol* 2015;96(1):143–55.
- [45] Kermani F, Gharavian A, Mollazadeh S, Kargozar S, Youssefi A, Vahdati Khaki J. Silicon-doped calcium phosphates; the critical effect of synthesis routes on the biological performance. *Mater Sci Eng C* 2020;111:110828.
- [46] Bohner M. Silicon-substituted calcium phosphates—a critical view. *Biomaterials* 2009;30(32):6403–6.
- [47] Vahabzadeh S, Roy M, Bose S. Effects of silicon on osteoclast cell mediated degradation, in vivo osteogenesis and vasculogenesis of brushite cement. *J Mater Chem B* 2015;3(46):8973–82.
- [48] Moses JC, Dey M, Bavya Devi K, Roy M, Nandi SK, Mandal BB, et al. Synergistic effects of silicon/zinc doped brushite and silk scaffolding in augmenting the osteogenic

- and angiogenic potential of composite biomimetic bone grafts. *ACS Biomater Sci Eng* 2019;5(3):1462–75.
- [49] Jacobs A, Renaudin G, Forestier C, Nedelec J-M, Descamps S. Biological properties of copper-doped biomaterials for orthopedic applications: a review of antibacterial, angiogenic and osteogenic aspects. *Acta Biomater* 2020.
- [50] Dong C, Feng W, Xu W, Yu L, Xiang H, Chen Y, et al. The copper age: copper (Cu)-involved nanotheranostics. *Adv Sci* 2020;7(21):2001549.
- [51] Tsai C-Y, Cameron Finley J, Ali SS, Patel HH, Howell SB. Copper influx transporter 1 is required for FGF, PDGF and EGF-induced MAPK signaling. *Biochem Pharmacol* 2012;84(8):1007–13.
- [52] Hui Y, Dong Z, Wenkun P, Yao D, Huichang G, Tongxiang L. Facile synthesis of copper doping hierarchical hollow porous hydroxyapatite beads by rapid gelling strategy. *Mater Sci Eng C* 2020;109:110531.
- [53] Elrayah A, Zhi W, Feng S, Al-Ezzi S, Lei H, Weng J. Preparation of micro/nano-structure copper-substituted hydroxyapatite scaffolds with improved angiogenesis capacity for bone regeneration. *Materials* 2018;11(9):1516.
- [54] Ai F, Chen A, Yan J, Yang K, Li S, Duan H, et al. Hydroxyapatite scaffolds containing copper for bone tissue engineering. *J Sol-Gel Sci Technol* 2020;95(1):168–79.
- [55] Guo C, Li L, Li S, Wang Y, Yu X. Preparation, characterization, bioactivity and degradation behavior in vitro of copper-doped calcium polyphosphate as a candidate material for bone tissue engineering. *RSC Adv*, 2017;7(67):42614–26.
- [56] Kargozar S, Lotfibakhshaiesh N, Ai J, Samadikuchaksaraie A, Hill RG, Shah PA, et al. Synthesis, physico-chemical and biological characterization of strontium and cobalt substituted bioactive glasses for bone tissue engineering. *J Non-Crystall Solids* 2016;449:133–40.
- [57] Ji Z, Yang G, Shahzidi S, Tkacz-Stachowska K, Suo Z, Nesland JM, et al. Induction of hypoxia-inducible factor-1 $\alpha$  overexpression by cobalt chloride enhances cellular resistance to photodynamic therapy. *Cancer Lett* 2006;244(2):182–9.
- [58] Birgani ZT, Fennema E, Gijbels MJ, Boer J, Blitterswijk C, Habibovic P. Stimulatory effect of cobalt ions incorporated into calcium phosphate coatings on neovascularization in an in vivo intramuscular model in goats. *Acta Biomater* 2016;36:267–76.
- [59] Zhang M, Wu C, Li H, Yuen J, Chang J, Xiao Y. Preparation, characterization and in vitro angiogenic capacity of cobalt substituted  $\beta$ -tricalcium phosphate ceramics. *J Mater Chem* 2012;22(40):21686–94.
- [60] Kulanthaivel S, Roy B, Agarwal T, Giri S, Pramanik K, Pal K, et al. Cobalt doped proangiogenic hydroxyapatite for bone tissue engineering application. *Mater Sci Eng C* 2016;58:648–58.
- [61] Fani N, Farokhi M, Azami M, Kamali A, Lotfi Bakhshaiesh N, Ebrahimi-Barough S, et al. Endothelial and osteoblast differentiation of adipose-derived mesenchymal stem cells using a cobalt-doped CaP/silk fibroin scaffold. *ACS Biomater Sci Eng* 2019;5(5):2134–46.
- [62] Moseman RF. Chemical disposition of boron in animals and humans. *Environ Health Persp*, 1994;102(suppl 7):113–17.
- [63] Bent-al-hoda Movahedi Najafabadi M, Abnosi H. Boron induces early matrix mineralization via calcium deposition and elevation of alkaline phosphatase activity in differentiated rat bone marrow mesenchymal stem cells. *Cell J (Yakhteh)* 2016;18(1):62.
- [64] Park M, Li Q, Shcheynikov N, Zeng W, Muallem S. NaBC1 is a ubiquitous electrogenic Na<sup>+</sup>-coupled borate transporter essential for cellular boron homeostasis and cell growth and proliferation. *Mol Cell* 2004;16(3):331–41.



- [65] Dzondo-Gadet M, Mayap-Nzietchueng R, Hess K, Nabet P, Belleville F, Dousset B. Action of boron at the molecular level. *Biol Trace Element Res* 2002;85(1):23–33.
- [66] Durand LAH, Vargas GE, Romero NM, Vera-Mesones R, Porto-López JM, Boccaccini AR, et al. Angiogenic effects of ionic dissolution products released from a boron-doped 45S5 bioactive glass. *J Mater Chem B* 2015;3(6):1142–8.
- [67] Balasubramanian P, Büttner T, Miguez Pacheco V, Boccaccini AR. Boron-containing bioactive glasses in bone and soft tissue engineering. *J Eur Ceram Soc* 2018;38(3):855–69.
- [68] Kolmas J, Velard F, Jaguszewska A, Lemaire F, Kerdjoudj H, Gangloff SC, et al. Substitution of strontium and boron into hydroxyapatite crystals: Effect on physicochemical properties and biocompatibility with human Wharton-Jelly stem cells. *Mater Sci Eng C*, 2017;79:638–46.
- [69] Pazarçeviren AE, Tezcaner A, Keskin D, Topsoy Kolukısa S, Sürdem S, Evis Z. Boron-doped biphasic hydroxyapatite/ $\beta$ -tricalcium phosphate for bone tissue engineering. *Biol Trace Element Res* 2021;199(3):968–80.
- [70] Tunçay EÖ, Demirtaş TT, Gümüşderelioğlu M. Microwave-induced production of boron-doped HAp (B-HAp) and B-HAp coated composite scaffolds. *J Trace Elements Med Biol* 2017;40:72–81.
- [71] Rau JV, Fosca M, Fadeeva IV, Kalay S, Culha M, Raucci MG, et al. Tricalcium phosphate cement supplemented with boron nitride nanotubes with enhanced biological properties. *Mater Sci Eng C* 2020;114:111044.
- [72] Tang Y-Q, Wang Q-Y, Ke Q-F, Zhang C-Q, Guanb J-J, Guo Y-P. Mineralization of ytterbium-doped hydroxyapatite nanorod arrays in magnetic chitosan scaffolds improves osteogenic and angiogenic abilities for bone defect healing. *Chem Eng J* 2020;387:124166.
- [73] Tan Z, Zhou B, Zheng J, Huang Y, Zeng H, Xue L, et al. Lithium and copper induce the osteogenesis-angiogenesis coupling of bone marrow mesenchymal stem cells via crosstalk between canonical Wnt and HIF-1 $\alpha$  signaling pathways. *Stem Cells Int* 2021;2021:1–15.
- [74] Augustine R, Dominic EA, Reju I, Kaimal B, Kalarikkal N, Thomas S. Investigation of angiogenesis and its mechanism using zinc oxide nanoparticle-loaded electrospun tissue engineering scaffolds. *RSC Adv* 2014;4(93):51528–36.
- [75] Barui AK, Nethi SK, Patra CR. Investigation of the role of nitric oxide driven angiogenesis by zinc oxide nanoflowers. *J Mater Chem B* 2017;5(18):3391–403.
- [76] Xiao D, Yang F, Zhao Q, Chen S, Shi F, Xiang X, et al. Fabrication of a Cu/Zn co-incorporated calcium phosphate scaffold-derived GDF-5 sustained release system with enhanced angiogenesis and osteogenesis properties. *RSC Adv* 2018; 8(52):29526–34.
- [77] Xiao D, Zhang J, Zhang C, Barbieri D, Yuan H, Moroni L, et al. The role of calcium phosphate surface structure in osteogenesis and the mechanisms involved. *Acta Biomater* 2020;106:22–33.
- [78] Piard C, Luthcke R, Kamaliddinov T, Fisher J. Sustained delivery of vascular endothelial growth factor from mesoporous calcium-deficient hydroxyapatite microparticles promotes in vitro angiogenesis and osteogenesis. *J Biomed Mater Res A* 2021;109(7): 1080–1087.
- [79] Haider A, Haider S, Han SS, Kang I-K. Recent advances in the synthesis, functionalization and biomedical applications of hydroxyapatite: a review. *RSC Adv* 2017;7(13): 7442–7458.

- [80] Lode A, Wolf-Brandstetter C, Reinstorf A, Bernhardt A, König U, Pompe W, et al. Calcium phosphate bone cements, functionalized with VEGF: release kinetics and biological activity. *J Biomed Mater Res A* 2007;81A(2):474–83.
- [81] Keefe AD, Pai S, Ellington A. Aptamers as therapeutics. *Nat Rev Drug Discov* 2010;9(7):537–50.
- [82] Son J, Kim J, Lee K, Hwang J, Choi Y, Seo Y, et al. DNA aptamer immobilized hydroxyapatite for enhancing angiogenesis and bone regeneration. *Acta Biomater* 2019;99:469–78.
- [83] Gaebler A, Schaefer T, Fischer K, Scharnweber D, Mauth C, Schwenzer B. Peptide linkers for the immobilization of bioactive molecules on biphasic calcium phosphate via a modular immobilization system. *Acta Biomater* 2013;9(1):4899–905.
- [84] Parent M, Baradari H, Champion E, Damia C, Viana-Trecant M. Design of calcium phosphate ceramics for drug delivery applications in bone diseases: A review of the parameters affecting the loading and release of the therapeutic substance. *J Control Release* 2017;252:1–17.
- [85] Xu Q, Tanaka Y, Czernuszka JT. Encapsulation and release of a hydrophobic drug from hydroxyapatite coated liposomes. *Biomaterials* 2007;28(16):2687–94.
- [86] Sukhodub L, Sukhodub LF, Kumeda MO, Prylutska SV, Deineka V, Prylutsky YL, et al. C60 fullerene loaded hydroxyapatite-chitosan beads as a promising system for prolonged drug release. *Carbohydr Polym* 2019;223:115067.
- [87] Kermanian M, Naghibi M, Sadighian S. One-pot hydrothermal synthesis of a magnetic hydroxyapatite nanocomposite for MR imaging and pH-Sensitive drug delivery applications. *Heliyon* 2020;6(9):e04928.
- [88] Chen R, Shi J, Zhu B, Zhang L, Cao S. Mesoporous hollow hydroxyapatite capped with smart polymer for multi-stimuli remotely controlled drug delivery. *Microporous Mesoporous Mater* 2020;306:110447.
- [89] Chang C-H, Liao T-C, Hsu Y-M, Fang H-W, Chen C-C, Lin F, et al. A poly (propylene fumarate)–Calcium phosphate based angiogenic injectable bone cement for femoral head osteonecrosis. *Biomaterials* 2010;31(14):4048–55.
- [90] Yu W-L, Sun T-W, Qi C, Zhao H-K, Ding Z-Y, Zhang Z-W, et al. Enhanced osteogenesis and angiogenesis by mesoporous hydroxyapatite microspheres-derived simvastatin sustained release system for superior bone regeneration. *Sci Rep* 2017;7(1):1–16.
- [91] Khurana K, Guillem-Marti J, Soldera F, Mücklich F, Canal C, Ginebra M-P. Injectable calcium phosphate foams for the delivery of Pitavastatin as osteogenic and angiogenic agent. *J Biomed Mater Res B Appl Biomater* 2020;108(3):760–70.
- [92] Elimelech R, Khoury N, Tamari T, Blumenfeld I, Gutmacher Z, Zigdon-Giladi H. Use of transforming growth factor- $\beta$  loaded onto  $\beta$ -tricalcium phosphate scaffold in a bone regeneration rat calvaria model. *Clin Implant Dentistry Relat Res* 2019;21(4):593–601.
- [93] Sun X, Kang Y, Bao J, Zhang Y, Yang Y, Zhou X. Modeling vascularized bone regeneration within a porous biodegradable CaP scaffold loaded with growth factors. *Biomaterials* 2013;34(21):4971–81.
- [94] Gbureck U, Hölzel T, Doillon CJ, Müller FA, Barralet JE. Direct printing of bioceramic implants with spatially localized angiogenic factors. *Adv Mater* 2007;19(6):795–800.
- [95] Wang C, Lai J, Li K, Zhu S, Lu B, Liu J, et al. Cryogenic 3D printing of dual-delivery scaffolds for improved bone regeneration with enhanced vascularization. *Bioactive Mater* 2021;6(1):137–45.

## Non-Print Items

### Abstract

Calcium phosphate (CaP) bioceramics represent versatile biomaterials in the biomedical setting, especially for managing damaged hard tissues. Hydroxyapatite (HAp) and tricalcium phosphate (TCP) are among the most well-known CaPs bioceramics with diverse applications in tissue engineering. These substances offer exceptional characteristics for bone repair and regeneration, including excellent biocompatibility, osteoconductivity, osteoinductivity, and osteogenesis. The ability of CaPs to induce new blood vessel formation, that is, angiogenesis, is considered an additional advantage for their in vivo applications. The chemical composition and physical properties (e.g., topography) of CaPs bioceramics were identified as key determinants in advancing angiogenesis. Current research emphasizes making modifications in the basic formulation of CaPs for enhancing their angiogenic capacity. Doping of specific types of metallic elements (e.g., copper) into CaPs structure can create more potent angiogenic biomaterials. In addition, surface functionalization of CaPs bioceramics by proangiogenic growth factors and other chemicals was proven to be effective in improving neovascularization and accelerated wound healing. It is worth noting that CaPs can also be utilized for loading and delivery of proangiogenic cargoes (growth factors, phytochemicals, etc.) with the aim of accelerated tissue healing. Along with technological progress, the development of proangiogenic CaPs scaffolds by using three-dimensional (3D) printing provides great opportunities for researchers and scientists who work on tissue engineering and regenerative medicine. In this chapter, we have tried to present and criticize the angiogenic capacity of different types of CaPs bioceramics and draw a conclusion based on their usability for accelerating bone tissue healing based on the current knowledge.

### Keywords

Calcium phosphates; Hydroxyapatite; Tricalcium phosphate (TCP); Angiogenesis; Neovascularization; Bone tissue engineering