



# Characteristics and Antibacterial Effect of Chitosan Coating Nanoemulsion Containing *Zataria multiflora* and *Bunium persicum* Essential Oils Against *Listeria monocytogenes*

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## Abstract

**Background:** Nowadays, finding natural compounds with antimicrobial properties against pathogens is very important, especially for the food and drug industries.

**Objectives:** The antibacterial activity of chitosan coatings nanoemulsion (NE) containing *Zataria multiflora* and *Bunium persicum* essential oils (EOs) was evaluated in a food model (chicken breast fillets) during 15 days of refrigerated storage.

**Methods:** The chicken breast fillets were divided into seven groups: control, chitosan 2%, sonicated chitosan 2%, chitosan NE coating containing *Z. multiflora* EO (ZMEO, 0.5%, and 1%) and chitosan NE coating containing *B. persicum* EO (BPEO, 0.5%, and 1%). Characteristics of chitosan NE coatings containing EOs were analyzed. Moreover, the antimicrobial activity of coatings against *Listeria monocytogenes* was investigated.

**Results:** The results showed good properties of the NE coatings. The analysis of EOs revealed that the major components for ZMEO were carvacrol (51.55%) and thymol (25.49%). In addition, the main components of BPEO were p-cumic aldehyde (38.39%) and p-cymene (18.36%). All treatments exhibited antimicrobial properties; however, the best result was recorded for chitosan NE coating containing 1% ZMEO, which was the lowest amount of *L. monocytogenes* (7.61 Log CFU/g). Moreover, *L. monocytogenes* analysis for chitosan NE coating containing 1% BPEO samples was 7.73 Log CFU/g.

**Conclusions:** Therefore, based on the results of this study, chitosan NE coating containing ZMEO and BPEO as natural preservatives can be recommended for meat products, especially chicken meats.

**Keywords:** *Listeria monocytogenes*, *Bunium persicum* Essential Oil, *Zataria multiflora* Essential Oil, Chitosan Edible Coating, Nanoemulsion, Chicken Breast Fillet

## 1. Background

Chicken meat is very susceptible to the growth of spoilage, pathogenic microorganisms, and oxidative reactions and has a very short shelf-life. The consumption of chicken meat has increased during last several years because of low-fat content, high nutritional value, distinctive flavor, and comparatively low cost of production compared to other meat products. High protein content, high moisture level, and relatively high pH value lead to microbial spoilage of meat, whereas aerobic condition causes the oxidation of lipid and protein (1, 2). There has been

an increasing demand for healthy and safe foods during the last decades since several serious diseases are caused by food-borne pathogenic microorganisms (3). Each year, 48 million individuals are estimated to get sick from food-borne diseases in the United States (4). The presence of microorganisms in foodstuffs has a critical significance, as they can decompose and deteriorate food quality and some health problems. Thermal processing not only could be effective in inactivating pathogenic bacteria in food, but on the other hand, it could lead to the deterioration of nutritional and organoleptic properties of the food (3, 5).

Among the various types of known food-borne diseases, *Salmonella* and *Listeria* are known as major pathogenic bacteria in raw meat (beef and chicken), eggs (4), raw vegetables (6), dairy products (7), and ready-to-eat foods (5).

Many consumers have become aware of chemical preservatives' adverse effects to retard microbial growth in meat products (8). This knowledge formed a trend toward green consumerism and the replacement of chemical preservatives with natural antimicrobial agents (1). Therefore, many efforts have been carried out to investigate edible and natural compounds with antimicrobial properties to increase the shelf life and quality of various foods.

Many approaches have been proposed to improve the shelf life of food (9); one of them is the application of edible coatings (10). Chitosan (a cationic polysaccharide) is one of the edible coatings applied in the food industry owing to its bioactive features, including antifungal, antibacterial, and antioxidative activities. It is a natural biopolymer obtained from crab and shrimp shells and has been classified as generally recognized as safe (GRAS) (11-13). Chitosan coatings exhibited antibacterial activity against gram-negative and gram-positive bacteria (14). Their antimicrobial activity can be improved via some approaches. For instance, chitosan coatings can be incorporated with essential oils (EOs). Essential oils such as *Bunium persicum* and *Zataria multiflora* are originated from herbal plants and well known as natural antimicrobial agents (15).

Nanoemulsions (NEs) are colloidal systems with an oil phase dispersed in an aqueous phase (16). Incorporating NEs with EOs leads to an increase in the contact surface of EOs. It reduces their necessary dose, which is beneficial since it reduces their undesirable effects on the organoleptic properties of foods (17, 18). Nanoemulsions containing EOs or their components, proved to be effective against a wide range of microorganisms (19, 20).

Chicken meat is a staple food in the human diet, but it is an enriched media for the growth of pathogenic microorganisms and spoilage bacteria. Many pathogenic bacteria can be found in chicken meats; among them, *Listeria* is of the major pathogenic bacteria in raw meat (beef and chicken), eggs (4), raw vegetables (6), dairy products (7), and ready-to-eat foods (5).

Few studies have used active chitosan coatings, especially NE forms to control pathogenic bacteria. Shukla *et al.* (21) applied chitosan coating combined with *Syzygium aromaticum* in chicken patties and improved the shelf life of samples. In another study, chitosan and pomegranate were used to improve the quality of chicken samples at 4°C (1). However, to our knowledge, no study has investigated the antibacterial effect of chitosan coating NE containing

*Zataria multiflora* essential oil (ZMEO) and *Bunium persicum* essential oil (PBEO) against pathogenic bacteria on chicken meat.

## 2. Objectives

The aim of the present study was to (1) prepare and evaluate the characteristic of chitosan coating NEs embedded with ZMEO and PBEO; (2) assess the antibacterial properties of NE against *Listeria monocytogenes* in chicken breast fillets during 15 days of storage at 4°C.

## 3. Methods

### 3.1. Materials

Extraction and analysis of *Z. multiflora* and *B. persicum* were carried out at the Iranian Institute of Medicinal Plants, Karaj, Alborz province, Iran. The EOs were kept at 4°C in the refrigerator away from direct light until the analysis time. All of the culture media were obtained from Merck (Darmstadt, Germany). *Listeria monocytogenes* (ATCC: 7644, 7834, 10671, 82119) strains were prepared from the Department of Food Hygiene, Faculty of Veterinary Medicine, Ferdowsi University of Mashhad, Mashhad, Iran.

### 3.2. Chitosan Coating Nanoemulsion Containing *Zataria multiflora* Essential Oil and *Bunium persicum* Essential Oil Preparation

Chitosan coating NE containing ZMEO and PBEO with some modifications was formulated according to Severino *et al.* (22). The non-ionic surfactant of Tween 80 (HLB = 15) and sterile distilled water were used to prepare the coating. Distilled water was used to dissolve chitosan (2% w/v); subsequently, acetic acid glacial (1% w/v) was added to the mixture. Then, a plasticizer (0.75 mL/g glycerol) was added to the mixture, and they were stirred for 10 min to obtain a transparent mixture.

After preparation of the chitosan solution, ZMEO and PBEO were dissolved in two concentrations (0.5% and 1%) with an emulsifier (Tween 80, 0.2% w/v) and stirred by using a hotplate magnetic stirrer (IKA, C-MAG HS 10, Germany) to acquire a uniform, stable, and transparent homogenous mixture. Then, it was transferred to a beaker containing chitosan solution. Chitosan solution containing EOs were homogenized using an Ultra Turrax T25 (IKA Labortechnik, Germany) high shear mixer at 3,000 rpm for 3 min and ultrasound probe sonicator device (50°C, pulse; 45 s and rest; 15 s) for 6 minutes, to form NE. The investigated treatments were chitosan 2%, sonicated chitosan 2%, chitosan coating NE containing ZMEO at two levels (0.5 and 1%) and chitosan coating NE containing PBEO at two levels (0.5 and 1%).

### 3.3. Droplet Size, Size Distribution, and $\zeta$ -potential and Preparation of *Listeria monocytogenes*

$\zeta$ -potential test shows the stability of emulsions. The analysis was carried out according to Noori et al. (11). The mean particle size (Z-average) was evaluated using the dynamic light scattering (DLS) at 25°C on a Zetasizer Nano S90, (Malvern Instruments, Malvern, Worcestershire, UK). The dilution was performed with distilled water (100: 1) before the measurement. All of the measurements were recorded at 25°C.

To prepare suspensions of *L. monocytogenes* (four strains), bacteria were cultured in 10 mL of BHI, and incubation was carried out at 37°C for 24 hours. Then, they were incubated for 18 hours at 37°C in supplementary culture media (BHI), and bacterial suspensions were prepared. Subsequently, bacterial suspensions were diluted to obtain 0.5 MacFarland turbidity ( $10^8 \times 1.5$  CFU/mL).

### 3.4. Chicken Fillets Preparation, Inoculation of the Bacteria, and *Listeria monocytogenes* Analysis

The chicken breasts were obtained from a poultry vendor in Mashhad, Iran during the summer of 2018. They were filleted and transported to the laboratory. Then, they were washed with cold tap water. From chicken breasts  $10 \pm 1$  g fillets were prepared. Then the samples were immersed in ethanol solution (70% v/v) and burnt to remove the surface microorganisms. In the end, 50  $\mu$ L of  $1.5 \times 10^8$  CFU/mL dilution of *L. monocytogenes* strains were inoculated to achieve a final concentration of  $\sim 10^6$  CFU/g. After inoculation of the bacteria on each sample, they were kept at room temperature for 30 to 45 minutes, to attach the bacteria into the surface of the samples. Then, the fillet samples were randomly divided into seven groups (as presented in Table 1), and were dipped in different coating treatments for 1 min and then drained for 30 min. Finally, the treated samples were placed at  $4 \pm 1^\circ\text{C}$  for 15 days and tested every three days (day 0, 3, 6, 9, 12, and 15).

Ninety mL sterile peptone water (0.1%) and 10 g of fillets were added in a plastic bag and homogenized by a stomacher (Seward Medical, London, UK) for three minutes. Decimal dilutions of homogenized samples were made, and 10  $\mu$ L of each dilution were transferred on Palcam agar and Xylose-lysin-decarboxycholelate-agar (XLD) to count *L. monocytogenes*. The incubation was done for 24–48 h at 37°C.

### 3.5. Statistical Analysis

The results were presented as means  $\pm$  standard deviations (SD). The results were analyzed through one-way analysis of variance (ANOVA). All analysis was carried out in triplicate, and the statistical analysis was done using

SPSS version 21 software. Analysis of variance and Bonferroni post hoc test or Dunnett T3 post-test was used to determine the significant difference at the level of  $P < 0.05$ . Also, to evaluate the stability of the treatments Kruskal-Wallis, Friedman, and Wilcoxon signed-rank tests were carried out.

## 4. Results and Discussion

### 4.1. *Zataria multiflora* and *Bunium persicum* Essential Oils Composition

The analysis of ZMEO and BMEO were performed by GC-MS, which showed the presence of 13 and 17 components, respectively (Table 2). Also, 91.44% and 94.54% of chemical compounds were identified in ZMEO and BMEO, respectively. The main components of ZMEO were thymol (25.49%), carvacrol (51.55%), alpha-terpinen (4.44%), and p-cymene (5.23%), while, in the case of BPEO p-cymene (18.36%), gamma terpinene (7.38%), cumene (4.69%), p-cumic aldehyde (38.39%), and 2-carene-10-al (13.26%) were the major constituents. The data obtained from this study was in accordance with another study from Iran (23). However, in another study, the main components of the BPEO were  $\alpha$ -methyl-benzenemethanol (14.59%), p-cuminaldehyde (23.50%),  $\gamma$ -terpinene (13.10%), and  $\beta$ -cymene (8.48%) that were different from the present results (24). These differences may be attributed to factors like plant growth phase, geographic location, and genetic and environmental factors (25).

### 4.2. Particle Size of Nanoemulsions

Table 3 shows the particle size for NEs as well as the polydispersity index. Based on the results, NEs were stable and showed a low particle diameter and uniform distribution. As represented in Table 2, the highest particle size of the chitosan coating combined with essential oils was about  $1363 \pm 5$  nm, and when converted to sonicated chitosan, the mean particle size was reduced to  $532 \pm 2$  nm. BPEO 0.5% and 1% particle size were  $159 \pm 1$  nm and  $188 \pm 1$  nm, respectively, indicating the lowest particle size. The lowest particle size was associated with chitosan containing BPEO 0.5% with a mean particle diameter of 159 nm, similar to the results of the study done by Artiga-Artigas et al. (26). In our study, the particle size was measured by DLS method, and the particle size was less than 600 nm in the range, which corresponded with the data obtained by the study of Jafari et al. (27), Hatanaka et al. (28) and Salvia-Trujillo et al. (29). In another study performed by Sow et al. (30), the average size of particles was  $309 \pm 19$  nm (mean  $\pm$  SD), which was in agreement with the present study, because the particle size was below 600 nm.

**Table 1.** List of the Treatments in the Present Study

No.	Treatment	Description
1	CON	Samples without any coating solution (control)
2	CH	Samples coated with chitosan solution
3	S + CH	Samples coated with sonicated-chitosan solution
4	NANO + CH + ZMEO 0.5%	Samples coated with chitosan solution containing <i>Zataria multiflora</i> essential oil (EO) 0.5% (w/v)
5	NANO + CH + ZMEO 1%	Samples coated with chitosan solution nanoemulsion (NE) containing <i>Z. multiflora</i> EO 1% (w/v)
6	NANO + CH + BPEO 0.5%	Samples coated with chitosan solution NE containing <i>Bunium persicum</i> EO 0.5% (w/v)
7	NANO + CH + BPEO 1%	Samples coated with chitosan solution NE containing <i>B. persicum</i> EO 1% (w/v)

**Table 2.** Chemical Composition of *Zataria multiflora* Essential Oil and *Bunium persicum* Essential Oil Analyzed by GC/MS

<i>Zataria multiflora</i> Essential Oil			<i>Bunium persicum</i> Essential Oil		
Compounds	RT	Relative Percent of Compounds	Compounds	RT	Relative Percent of Compounds
1R- $\alpha$ -Pinene	11.04	1.68	Cumene	11.937	4.69
- $\alpha$ -Phellandrene	13.66	0.1	Beta- Pinene	14.355	3.89
(+)-4-Carane	14.11	1.12	(+)-Camphene	14.975	0.50
p-Cymene	14.43	5.23	Alpha-Terpinen	15.478	0.36
a-Terpinen	15.71	4.44	P-Cymene	16.059	18.36
Bornel	19.96	0.11	Gama-Terpinen	17.222	7.38
Terpinen-4-ol	20.37	0.39	R-(+)-Pulegone	22.739	2.73
Alpha Terpineol	20.98	0.21	Alpha-Terpieol	23.560	0.34
Bornyl acetate	24.50	0.20	3-Caren-10-al	24.033	0.14
Thymol	25.60	25.49	P-Cumic aldehyde	25.737	38.39
Carvacrol	26.07	51.55	Alpha-Thujenal	26.171	1.27
Valencen	31.91	0.60	Phellandral	26.964	0.35
Caryophyllene oxide	39.23	0.32	Isogeraniol	27.070	0.59
-	-	91.44%	2-Caren-10-al	27.442	13.26
			P-Cymen-7-ol	28.984	1.33
			Carvacrol	30.557	0.91
			Caryophyllen	35.307	0.05
			-	-	94.54%

Abbreviation: RT, retention time.

#### 4.3. Antimicrobial Effects of Chitosan Coating Nanoemulsion in Combination with *Zataria multiflora* and *Bunium persicum* Essential Oils

The effects of chitosan coating NE containing *Z. multiflora* and *B. persicum* EOs and its effects against *L. monocytogenes* inoculated on chicken breast fillets were investigated during 15 days of storage (4°C). The results are summarized in Tables 3 and 4.

Changes in bacterial counts during 15 days of storage were represented in Figure 1. Chitosan coating nanoemulsion containing ZMEO 1% showed the highest an-

timicrobial activity against *L. monocytogenes*, while simple chitosan coating represented low antimicrobial activity against the tested bacterium. The data indicated that after 15 days of storage, the control group showed an increase in *L. monocytogenes* (from 6.20 Log CFU/g on day 1 to 9.20 Log CFU/g on day 15). Treatment groups exhibited a microbial load of 6.07 Log CFU/g up to 9 Log CFU/g, indicating a slight decrease compared to the control group. This agreed with the result of Hassanzadeh et al. (12). The antimicrobial activity of chitosan coatings can be found in many researches, reporting similar results (14, 31, 32). An-

**Table 3.** Results of Particle Size, Particle Size Distribution and zeta  $\zeta$ -potential<sup>a, b</sup>

Treatment	Particle Size	$\zeta$ -potential	Particle Size Distribution	P Value
CH	1363 ± 5 nm <sup>A</sup>	53.566 ± 51 mV <sup>A</sup>	0.47 ± 0.015 <sup>A</sup>	< 0.05
S + CH	532 ± 2 nm <sup>B</sup>	43.13 ± 32 mV <sup>B</sup>	0.28 ± 0.015 <sup>B</sup>	
Nano + CH + ZMEO %0.5	243 ± 4 nm <sup>C</sup>	44.46 ± 3.5 mV <sup>C</sup>	0.30 ± 0.04 <sup>C</sup>	
Nano + CH + ZMEO %1	277 ± 2 nm <sup>D</sup>	37.46 ± 2.8 mV <sup>D</sup>	0.49 ± 0.02 <sup>D</sup>	
Nano + CH + BPEO %0.5	159 ± 1 nm <sup>E</sup>	47.33 ± 2.5 mV <sup>E</sup>	0.29 ± 0.02 <sup>E</sup>	
Nano + CH + BPEO %1	188 ± 1 nm <sup>F</sup>	35.86 ± 3.3 mV <sup>F</sup>	0.41 ± 0.02 <sup>F</sup>	

<sup>a</sup> Values are expressed as mean ± standard deviation.

<sup>b</sup> Different upper case letters in the same column indicate statistically significant differences among (P < 0.05).

**Table 4.** Reduction of *Listeria monocytogenes* Counts in Treatment Groups During 15 Days Storage at 4°C<sup>a</sup>

Mean Difference I - J	Group(J)	Chitosan	Nanochitosan	Nano-chitosan + BPEO 0.5%	Nano-chitosan + BPEO 1%	Nano-chitosan + ZMEO 0.5%	Nano-chitosan + ZMEO 1%
Group(I)							
Control		0.14*	0.35*	0.61*	1.03*	1.00*	1.25*
Chitosan			0.20*	0.47*	0.88*	0.86*	1.10*
Nanochitosan				0.26*	0.67*	0.65*	0.90*
Nano-chitosan + BPEO 0.5%					0.41*	0.02*	0.22*
Nano-chitosan + BPEO 1%						0.02	0.22*
Nano-chitosan + ZMEO 0.5%							0.24*

Abbreviations: BPEO, *Bunium persicum* essential oil; ZMEO, *Zataria multiflora* essential oil.

<sup>a</sup> \*Indicate a statistically significant difference (P < 0.05).

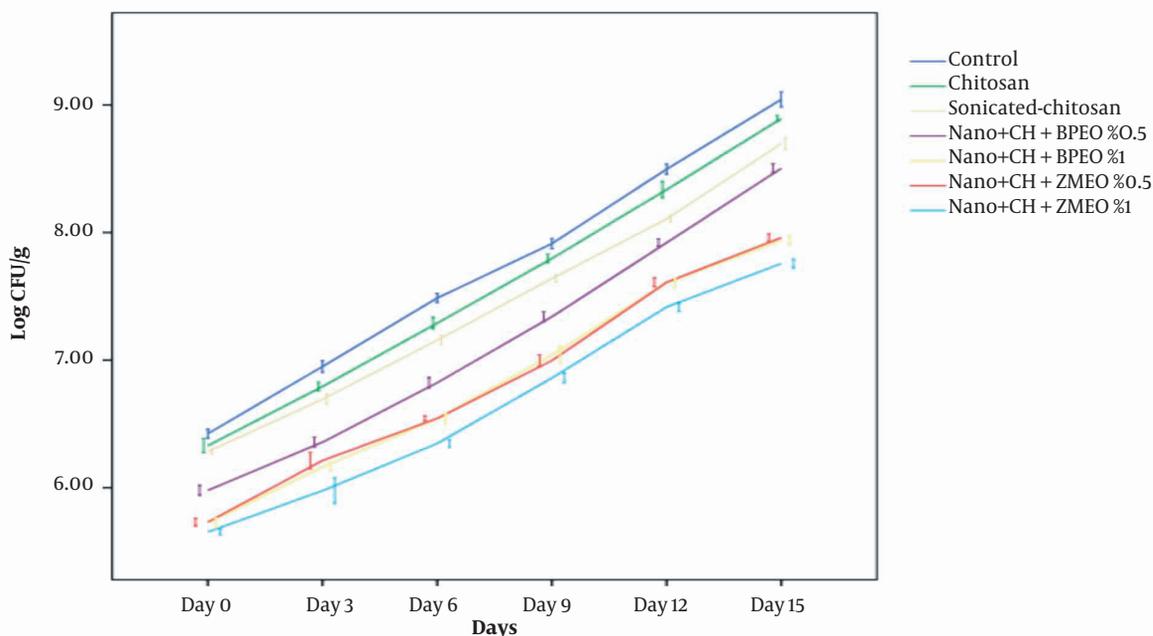
timicrobial properties of chitosan coatings may be associated with the alteration in the penetrability of the cell membrane, the interactions between the chitosan amine groups, and the electronegative load at the cell surface, which causes leakage of intracellular electrolytes and proteinaceous components (3, 33).

Sonicated-chitosan coating showed an increase of *L. monocytogenes* count from 5.89 Log CFU/g up to 8.76 Log CFU/g after 15 days of storage in the refrigerator, which showed a significant reduction in bacterial count in comparison with the chitosan and the control group. The reason is that in this form, the surface/volume ratio increases, and chitosan coating can easily penetrate to the bacterial membrane.

In the chitosan coating NE containing BPEO 0.5% and 1%, *L. monocytogenes* population decreased significantly from an initial value of 5.78 - 8.27 Log CFU/g to 5.41 Log CFU/g - 7.73 Log CFU/g after 15 days storage at the refrigerator compared to other treatments (nanochitosan, chitosan, and control groups). Antimicrobial activity of BPEO 1% was higher than BPEO 0.5% due to its higher concentration of EO. As mentioned above, chitosan coating alone has little antimicrobial activity. Noori et al. reported that adding EO to the NE system caused a significant increase in the an-

timicrobial activity (P < 0.05) (11). Antimicrobial activity of BEO is due to the presence of p-cymene, gamma terpinene, cumene, p-cumic aldehyde, and other active constituents, which are often terpenic compounds (24, 34). Topuz et al. reported that nanoemulsion of anise oil reduced *Escherichia coli* O157: H7 and *L. monocytogenes* population more than coarse emulsions (15). This could be due to low particle size and higher exposed area to the microbial cells (15).

In the NE chitosan coating containing ZMEO, 0.5% and 1% *L. monocytogenes* population increased significantly from 5.41 Log CFU/g - 7.83 Log CFU/g to 5.19 Log CFU/g - 7.61 Log CFU/g during 15 days of refrigerated storage. Although bacterial number has been increased during storage, ZMEO treatments (0.5% and 1%) significantly reduced bacterial count (1 and 1.25 Log CFU/g, respectively) compared to the control group. Among these treatments, ZMEO 1% had the highest antimicrobial activity against both bacteria due to the higher concentration of EO and thymol and carvacrol as the main antimicrobial compounds (34-36). These results agreed with those reported by Abdollahzadeh et al., indicating that the combination of thyme EO 0.4% and nisin with 1000 IU/g concentration were efficient means of *L. monocytogenes* controlling in minced fish (36). Moreover, in another study, *Z. multi-*



**Figure 1.** Changes in bacterial count (CFU/g) of chicken breast fillet samples inoculated with *L. monocytogenes* during the storage period (mean  $\pm$  SD)

flora which contained 36.62% carvacrol, was used in fresh chicken fillets and significantly improved the shelf life of samples (37).

Based on our findings (Figure 1), the antimicrobial activity of NE chitosan coating containing EOs was excellent against *L. monocytogenes*. All treatments exhibited an increasing trend during 15 days of storage at 4°C. However, the best result was observed in chitosan nanoemulsion combined with 1% ZMEO followed by chitosan nanoemulsion combined with 1% BPEO.

#### 4.4. Conclusions

In the current study, preparation, characteristic properties and application of nanochitosan-based edible coating containing ZMEO and BPEO were performed. The best results of nanoemulsion stability tests were related to chitosan coating containing nanoemulsion of PBE0 0.5%. The use of ZMEO and BPEO antimicrobial agents in nanochitosan coating can effectively maintain their antibacterial activity in chicken breast fillets during storage and can significantly reduce the growth of these two pathogens. Our results also showed that nanochitosan containing ZMEO 1% had the best antimicrobial effect on the growth of *L. monocytogenes* in chicken breast fillets. Finally, it should be considered that the introduced treatments cannot guarantee the safety of cold chicken breasts when contaminated with high doses (106 CFU/g) of *L. monocytogenes* due to their

ability to grow at cold temperatures. However, the samples treated with essential oils can reduce the microbial count for about 1.25 Log CFU/g after 15 days of storage at 4°C. Therefore, due to the interest of consumers and manufacturers in using natural additives instead of chemical additives in food, nanochitosan containing EOs can be used efficiently in chicken breast fillets to enhance its safety against the two pathogenic bacteria.

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#### Footnotes

**Authors' Contribution:** M. H.: Study concept and design, analysis and interpretation of data, critical revision of the manuscript for important intellectual content, study supervision, approved the final manuscript; A. A.: Study concept and design, drafting of the manuscript, critical revision of the manuscript for important intellectual content, approved the final manuscript; Z. B. Z.: Acquisition of data, drafting of the manuscript, approved the final manuscript;

S. K.: Acquisition of data, drafting of the manuscript, approved the final manuscript; M. R.: Acquisition of data, Statistical analysis, approved the final manuscript; S. M. A. N.: Analysis and interpretation of data, Statistical analysis, approved the final manuscript.

**Conflict of Interests:** The authors declare no conflict of interests. Only one of the co-authors (Seyyed Mohammad Ali Noori) is affiliated with the Jundishapur University of Medical Sciences. However, he is not a member of the editorial board of the journal nor does he have any role in the review process of the current manuscript.

**Data Reproducibility:** The data presented in this study are openly available in one of the repositories or will be available on request from the corresponding author by this journal representative at any time during submission or after publication. Otherwise, all consequences of possible withdrawal or future retraction will be with the corresponding author.

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